

Manuscript Number:

Title: Applications of ion mobility mass spectrometry for high throughput, high resolution glycan analysis

Article Type: SI: Glycans

Keywords: glycan analysis
glycan sequencing
mass spectrometry
ion mobility spectrometry
stereochemical configuration
glycoconjugates

Corresponding Author: Dr. Sabine L. Flitsch,

Corresponding Author's Institution: Univ. of Edinburgh

First Author: Sabine L. Flitsch

Order of Authors: Sabine L. Flitsch

Abstract: Abstract

Background

Diverse varieties of often heterogenous glycans are ubiquitous in Nature. They play critical roles in recognition events, act as energy stores and provide structural stability at both molecular and cellular levels. Technologies capable of fully elucidating the structures of glycans are far behind the other '-omic' fields. Liquid chromatography (LC) and mass spectrometry (MS) are currently the most useful techniques for high-throughput analysis of glycans. However, these techniques do not provide full unambiguous structural information and instead the gap in full sequence assignment is frequently filled by a priori knowledge of the biosynthetic pathways and the assumption that these pathways are highly conserved.

Scope of the review

This comprehensive review details the rise of the emerging analytical technique ion mobility spectrometry (IMS) (coupled to MS) to facilitate the determination of three-dimensional shape: the separation and characterization of isobaric glycans, glyco(peptides/proteins), glycolipids, glycosaminoglycans and other polysaccharides; localization of sites of glycosylation; or interpretation of the conformational change to proteins upon glycan binding.

Major conclusions

IMS is a highly promising new analytical route, able to provide rapid isomeric separation (ms timescale) of either precursor or product ions facilitating MS characterization. This additional separation also enables the deconvolution of carbohydrate MS(/MS) information from contaminating ions, improving sensitivity and reducing chemical noise. Derivation of collision cross sections (CCS) from IM-MS(/MS) data and subsequent calculations validate putative structures of carbohydrates from ab initio derived candidates. IM-MS has demonstrated that amounts of specific

glycan isomers vary between disease states, which would be challenging to detect using standard analytical approaches.

General significance

IM-MS is a promising technique that fills an important gap within the Glycomics toolbox, namely identifying and differentiating the three-dimensional structure of chemically similar carbohydrates and glycoconjugates.



The University of Manchester
Manchester Institute of Biotechnology

Prof. Sabine L. Flitsch
School of Chemistry &
Manchester Institute of Biotechnology
The University of Manchester
131 Princess Street
Manchester
M1 7DN

tel +44 (0) 161 306 5172

1st December 2015

Tiffany J Hicks
Managing Editor- Special Issues
BBA - General Subjects

Dear Editor

Please find attached the manuscript entitled

Applications of ion mobility mass spectrometry for high throughput, high resolution glycan analysis

which we wish to submit as review to the Special Issue BBA –General Subjects on ‘Glycans in personalized medicine’.

Kind regards,

Sabine L. Flitsch

Applications of ion mobility mass spectrometry for high throughput, high resolution glycan analysis

C.J. Gray,^{1*} B. Thomas,^{1*} R. Upton,^{1*} L.G. Migas,¹ C.E. Evers,² P.E. Barran^{1,3} and S.L. Flitsch¹

¹School of Chemistry & Manchester Institute of Biotechnology, The University of Manchester, 131 Princess Street, Manchester, M1 7DN, United Kingdom

²Department of Biochemistry, Institute of Integrative Biology, University of Liverpool, Crown Street, Liverpool, L69 7ZB, United Kingdom

³Michael Barber Centre for Collaborative Mass Spectrometry, Manchester Institute of Biotechnology, The University of Manchester, 131 Princess Street, Manchester, M1 7DN, United Kingdom

*Denotes authors corresponded equally

Abstract

Background

Diverse varieties of often heterogenous glycans are ubiquitous in Nature. They play critical roles in recognition events, act as energy stores and provide structural stability at both molecular and cellular levels. Technologies capable of fully elucidating the structures of glycans are far behind the other ‘-omic’ fields. Liquid chromatography (LC) and mass spectrometry (MS) are currently the most useful techniques for high-throughput analysis of glycans. However, these techniques do not provide full unambiguous structural information and instead the gap in full sequence assignment is frequently filled by *a priori* knowledge of the biosynthetic pathways and the assumption that these pathways are highly conserved.

Scope of the review

This comprehensive review details the rise of the emerging analytical technique ion mobility spectrometry (IMS) (coupled to MS) to facilitate the determination of three-dimensional shape: the separation and characterization of isobaric glycans, glyco(peptides/proteins), glycolipids, glycosaminoglycans and other polysaccharides; localization of sites of glycosylation; or interpretation of the conformational change to proteins upon glycan binding.

Major conclusions

IMS is a highly promising new analytical route, able to provide rapid isomeric separation (ms timescale) of either precursor or product ions facilitating MS characterization. This additional separation also enables the deconvolution of carbohydrate MS(/MS) information from contaminating ions, improving sensitivity and reducing chemical noise. Derivation of collision cross sections (CCS) from IM-MS(/MS) data and subsequent calculations validate putative structures of carbohydrates from *ab initio* derived candidates. IM-MS has demonstrated that amounts of specific glycan isomers

vary between disease states, which would be challenging to detect using standard analytical approaches.

General significance

IM-MS is a promising technique that fills an important gap within the Glycomics toolbox, namely identifying and differentiating the three-dimensional structure of chemically similar carbohydrates and glycoconjugates.

1. Introduction

Glycans and their conjugates (proteins, lipids, various anabolites) are found attached to the surface of cells, within cells and in the extracellular matrix. They are integral to an extraordinarily diverse range of biological processes including regulation of cell-cell recognition^[1], cell adhesion^[2] (including host-pathogen interactions^[3]), immune response,^[4] fertilization,^[3a, 5] trafficking, and intra- and extracellular signalling events. Even subtle changes in carbohydrate structure can result in vastly different interactions, make them susceptible to enzymatic hydrolysis or alter glycoconjugate tertiary structures and thus affect their observed biological response.^[2a, 6] Interestingly both the identity and levels (or presence) of specific glycan structures within individuals depends on numerous factors including gender,^[5a, 7] blood group,^[8] age,^[9] disease state^[6b, 6g, 10] and diet.^[11] It remains unclear *a priori* how changes in glycan structure will affect the resultant biological function.^[12] Given their ubiquity in nature, it is unsurprising that aberrant glycan structures have been identified as biomarkers for several disease states including various cancers,^[6a, 6d, 10c, 13] hereditary disorders,^[6g, 14] acute pancreatitis,^[10a] immune and cardiovascular deficiencies,^[6b, 12, 15] Alzheimer's disease^[16] and muscular dystrophy^[2a].

The incredible amount of three dimensional stereochemical information contained within glycans is biologically very important, as it is required to accommodate for their diverse functionalities even from a relatively small pool of monosaccharide building blocks. This complex 'chemical information' arises from: (1) different monosaccharide building blocks; (2) the potential to link these monomers at a number of different positions in two different stereochemistries, namely α - and β -, forming both linear and branched structures; (3) the ability for identical monosaccharides to have been incorporated with different ring connectivities, *i.e.* pyranose and furanose structures (4) further decoration with chemically diverse functional groups (*e.g.* phosphate, sulphate, methyl groups) (Figure 1). To understand and exploit glycan-driven functions, it is necessary to be able to unambiguously elucidate carbohydrate structures, in the absence of *a priori* information to be able to define structure-function relationships. To develop and exploit differences in glycan structures as potential biomarkers of disease, it will be necessary to define these structures in a high throughput manner to allow characterization on a person-to-person basis, permitting the development of novel

therapeutics or diagnostics.^[17] The demand for three-dimensional structure determination has resulted in fewer advances in comprehensive structural elucidation of glycans compared with proteins and nucleic acids, especially since glycan structures are not directly encoded by genetic information. Moreover, microheterogeneity, the low amounts available for analysis, and the chemical similarity between monosaccharide building blocks, which are often simple epimers of one another, hinder their full structural elucidation.

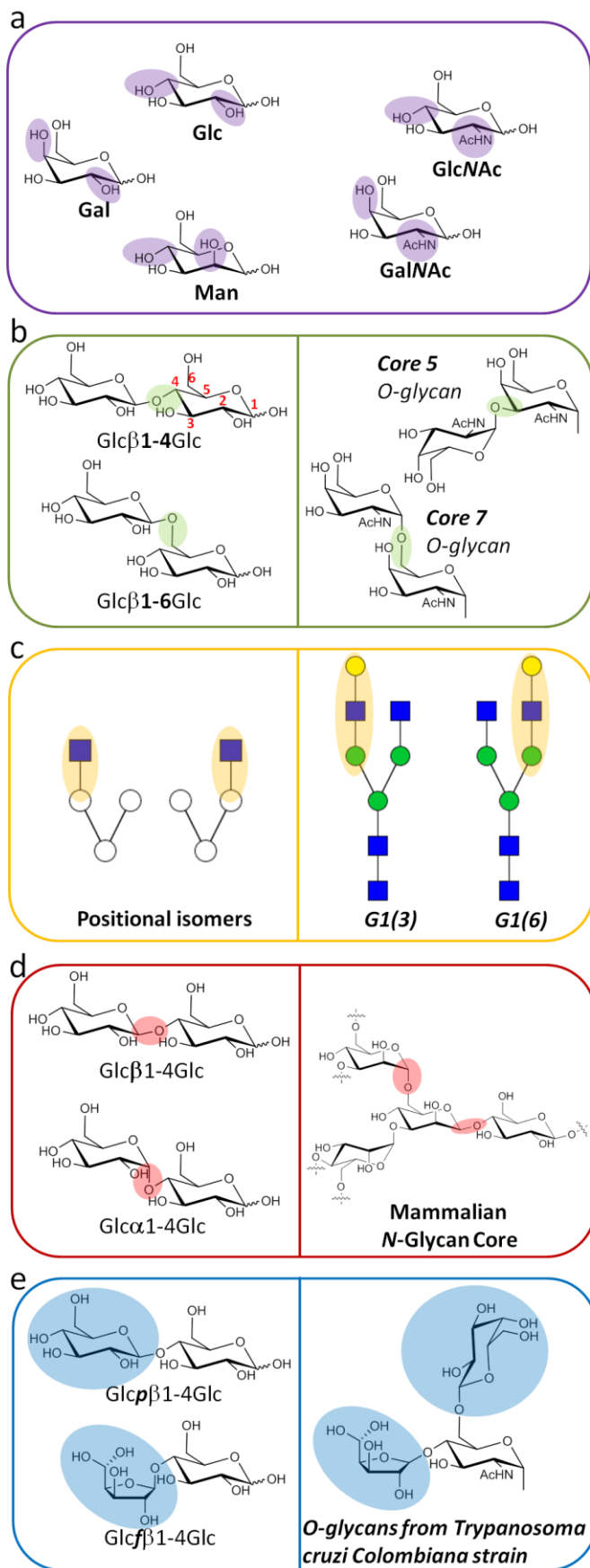


Figure 1 – Scheme depicting the various types of isomerism routinely observed in nature. The left side of the panel highlights the types of isomerism observed and the right side displays naturally occurring examples. Different stereochemistries associated with the monosaccharide building blocks (a) Regioisomers of glycosidic bond formation between two monosaccharide building blocks, which includes the potential formation of branched structures; (b) Positional isomers resulting from addition of a monosaccharide unit to different branching arms; (c) Anomeric glycosidic bond isomers α - and β -; (d) Finally, ring size isomers pyranose, *p*, and furanose, *f*, (e).

The current ‘gold standard’ for glycan analysis typically involves liquid chromatography (LC) separation,^[6a, 6b, 12, 18] which can also be coupled with the speed and sensitivity of tandem mass spectrometry (MS²) (Figure 2).^[19] Glycans are then characterized based on either their retention time against a standard and/or by their MS² spectrum.^[18b, 20] However, LC separation can require multiple different columns to gain significant separation of chemically similar glycan species and as a result it is often low throughput. Also, without the benefit of (synthetic) reference standards, LC provides no structural *de novo* information. MS alone is limited to characterising the monosaccharide class (*i.e.* hexose, N-acetylhexosamine, deoxy hexose *etc.*) and is insufficient to directly identify these monosaccharide units from each other without the use of an orthogonal sequencing approach. Given the intrinsic ability of carbohydrates to be isomeric, these strategies only often work for systems whose biosynthetic pathways are well understood (e.g. N-glycans) to facilitate in their identification. The inability of this analysis strategy to separate and characterize these iso(mer/bar)ic species from complex biological mixtures can hinder the development of specific glycomic approaches for assessing disease state.^[10d]

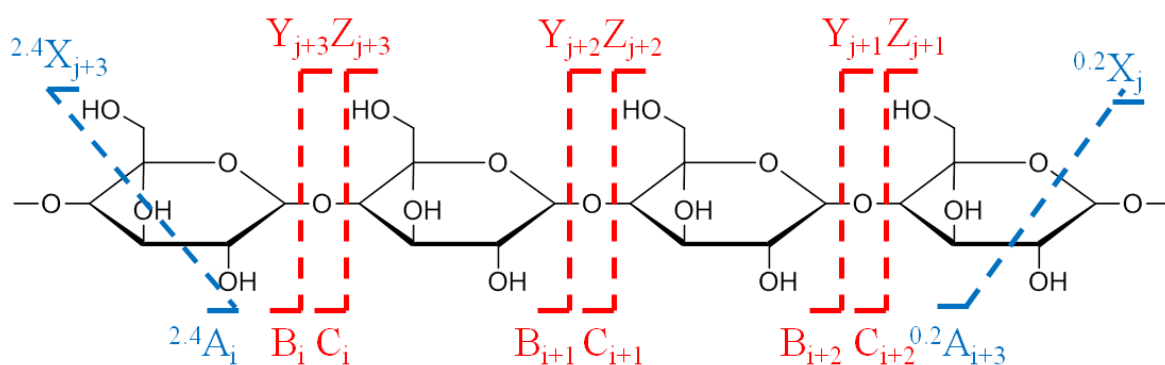


Figure 2 - Scheme depicting Domon-Costello nomenclature for carbohydrate product ions formed as a result of tandem mass spectrometry. Red and blue ions correspond to those formed as a result of fragmenting the glycosidic bond and the monosaccharide ring respectively.^[21]

Recently there has been a surge in the application of Ion Mobility-Mass Spectrometry (IM-MS) to enhance structural characterization and separation of glycans and glycoconjugates. Ion mobility spectrometry (IMS) is an analytical technique that measures the mobility, *K*, of gas-phase ions under the influence of an electric field in the presence of a buffer gas; the mobility is based on the size, shape and charge of the ions,^[22] analogous to electrophoresis in the condensed phase, although the timescale is much shorter (high- μ s to ms). Ions can be generated by a range of techniques, although for most IMS applications, ions are generated by ESI. This technique has been used for a variety of types of analyses, and has major applications in security and defence for example

in airports where it is routinely used to screen against explosives, chemical-warfare agents and drugs. The recent surge in use of ion mobility to analyse glycans is primarily as a result of the commercialization of hybrid IM-MS instrumentation (Synapt HDMS, 2006) and its potential to overcome challenges associated with analysis of glycans; namely separation of often isomeric carbohydrates and structural characterization of carbohydrates in conjugation with molecular dynamics (MD) simulations.

This review assesses the applicability of IM-MS towards biologically relevant glycans and the wealth of additional information that can be obtained from this high-throughput strategy. This will undoubtedly improve our understanding of glycan functions and thus could provide promising tools for further medicinal applications. Initially, the various IMS strategies routinely coupled to MS will be briefly discussed to help the reader appreciate the limitations and advantages of each of the techniques; however for a more thorough background into IMS the reader is encouraged to read excellent previous reviews.^[22b, 23] Similarly, computational aspects (such as MD simulations) that facilitate elucidation of gas-phase structures are briefly discussed, however, they are more extensively detailed elsewhere.^[24]

2. Ion mobility spectrometry

A number of unique ion mobility analysers have been coupled to MS, each with their own benefits and limitations (Figure 3). Each of these analysers are discussed in further detail.

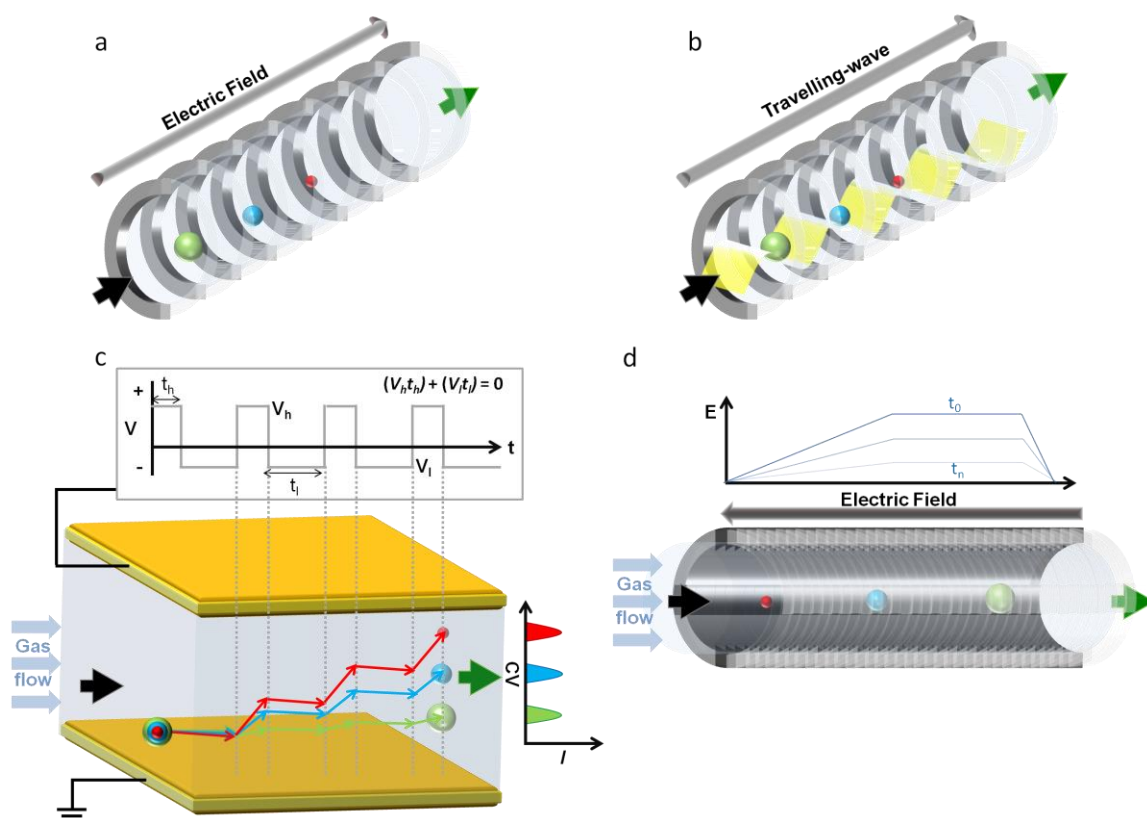


Figure 3 – Four different mobility cells for drift tube (a), travelling-wave (b), (high) field asymmetric (c) and trapped (d) ion mobility spectrometry. Ions of increasing collision cross section (Ω)-to-charge ratio are depicted as red, blue and green spheres. Solid black and green arrows correspond to where the ions enter and exit respectively.

2.1 Drift tube (DT)IMS

DTIMS is conceptually the simplest form of IMS. Ions are introduced (gated) into a cell consisting of a series of stacked electrodes filled with a static drift/buffer gas. A weak, uniform electric field ($\sim 5\text{-}15$ V/cm) is applied across the cell causing the ion packet to move through the cell. Ions with larger rotationally averaged collision cross section (CCS) undergo more collision/interaction events with the drift/buffer gas and as a result take longer to traverse the cell. This CCS represents the effective area of the gas-phase ion, under the precise set of experimental conditions that can interact with the drift gas, averaged over all orientations and thus can be related to structural or conformational features. Mobility separated ions are then separated by their m/z using a mass analyser that tends to be, but not exclusively, a time-of-flight device before being detected. The mobility or reduced mobility, K_0 , for a standard set of conditions can then be calculated for the ions using equations 1 and 2:

$$K = \frac{L}{t_d E} \quad (1)$$

$$K_0 = K \frac{P}{760} \frac{273.2}{T} = \frac{L}{t_d E} \frac{P}{760} \frac{273.2}{T} \quad (2)$$

Where L is the length of the drift cell, t_d is the drift time, E is the electric field, P is the pressure within the cell and T is the temperature of the buffer gas within the cell. K_0 can then be equated to the rotationally averaged CCS under a specific set of instrument conditions, Ω , according to the Mason-Schamp equation (3):^[25]

$$\Omega = \frac{1}{K_0} \frac{(18\pi)^{1/2}}{16} \frac{ze}{(k_b T)^{1/2}} \left[\frac{1}{m_I} + \frac{1}{m_B} \right]^{1/2} \frac{1}{N} \quad (3)$$

Where k_b is the Boltzmann constant, m_I is the mass of the ion, m_B is the mass of the buffer gas, z is the charge state, e is the elementary charge, and N is the number density of the drift gas; this relationship only holds true for low E/N (< 2 Td).^{[26] [27]} At high E/N the electric field causes ion heating and therefore a non-linear relationship between $1/K_0$ and Ω .

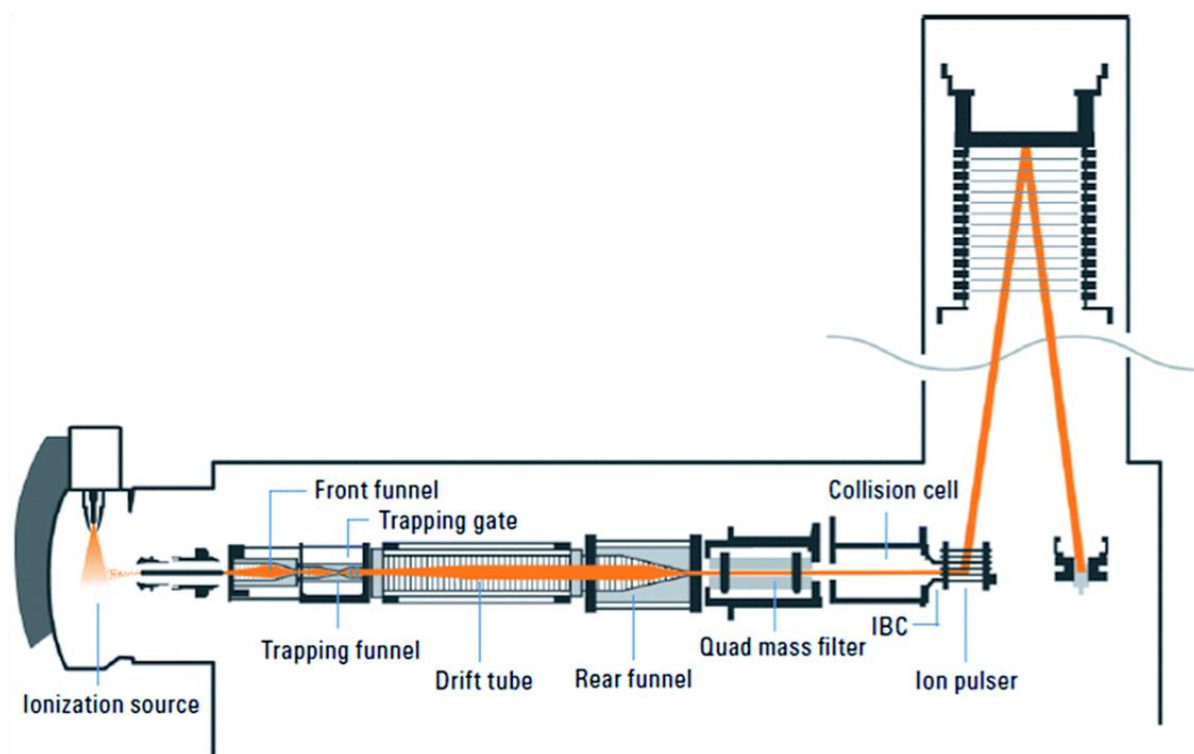


Figure 4 – Schematic of the commercially available Agilent DTIMS instrument.

The resolving power of IMS instrumentation is highly dependent on the choice of buffer gas. Classically for gas-phase structural (and separation) analysis, the buffer gas is an inert gas such as helium or nitrogen; these two gases have been extensively studied and permit determination of the CCS of ions of interest. In the past, other gases have been shown to offer superior separation capabilities, particularly gases that are polarizable (i.e. CO₂, SF₆ and N₂O). These gases allow for the improved separation of isomeric species, but this separation is highly dependent on the physical properties of the analyte.^[25b] The resolving power of modern IMS instruments tends to be in the order of 50-100 ($\Omega/\Delta\Omega$) and the increase in resolution tends to come at the expense of sensitivity. One major disadvantage of DTIMS is its low duty cycle. The DTIMS instrument has only been used by a handful of specialized research groups due to the lack of commercially available instrumentation, however, with the recent release of a commercial instrument (Agilent 6560 Ion Mobility Q-ToF; Figure 4), adoption of DTIMS and its application is expected to rise.^[28]

2.2 Travelling wave (TW)IMS

TWIMS was first commercialized by the release of the Synapt HDMS (2006, Waters; Figure 5).^[29] Since then several new improved iterations of the Synapt HDMS series and other instrumentation possessing TWIMS (i.e. Waters, Vion IMS QToF) have been released.^[30] Ions are guided through the mobility cell using a series of ring electrodes (stacked ring ion guide), in the presence of a static buffer gas (commonly N₂) akin to DTIMS. Travelling waves are generated by

raising the voltage on selected, periodically spaced electrodes and then moving the potential along the ring electrodes. Ions with smaller CCS undergo fewer collisions with the drift gas and ‘surf’ the waves, whereas larger ions undergo more collisions and are more likely to tumble over the back of the waves. Simultaneously a RF potential is applied to the electrodes creating a well at the centre of the cell, radially confining ions and preventing them from diffusing to the electrodes where they would annihilate. Mobility separation can be tuned by altering the wave height, velocity or the pressure of the mobility cell. Like DTIMS, TWIMS operates in the low E/N regime meaning CCS can be derived. However, ion trajectories through the mobility cell have not been fully elucidated and as a result the CCS cannot be directly measured from the drift time of the analyte. As a result, CCS derivation by TWIMS is achieved by calibration against a series of molecules whose CCS are known and ideally are of similar ‘class’ to the analyte being studied.^[31] When multiple charge states are present in either the calibrant or analyte, their CCS is often reported as a mass and charge state normalized value (Ω') calculated using the following equation (4):

$$\Omega' = \frac{\Omega(\mu)^{1/2}}{z} \quad (4)$$

where μ is the reduced mass. However, using Ω' values for calibration have been reported to introduce significant errors.^[31a] Unfortunately for structural applications, TWIMS data is commonly recorded in nitrogen and DTIMS in helium. TWIMS cannot operate with (pure) helium as a drift gas since the cell operates at a potential and pressure beyond the electrical breakdown limit of helium. As a result $^{TW}CCS_{N_2}$ is often converted to a representative $^{TW}CCS_{He}$ value for calibration purposes, although this is also reported to introduce error.^[31b, 32] The T-wave cell is also reported to produce significant ion heating, raising questions on the ability of the instrument to yield structural data representative of the solution phase (“native”). However, recent advances of the instrument design have been demonstrated to significantly reduce ion heating, with evidence that solution-phase protein, peptide and glycan conformers can be retained in the gas-phase.^[33] Despite these concerns, the ion heating is insufficient to break the covalent bonds, therefore TWIMS is a suitable technique for structural analysis of carbohydrates.

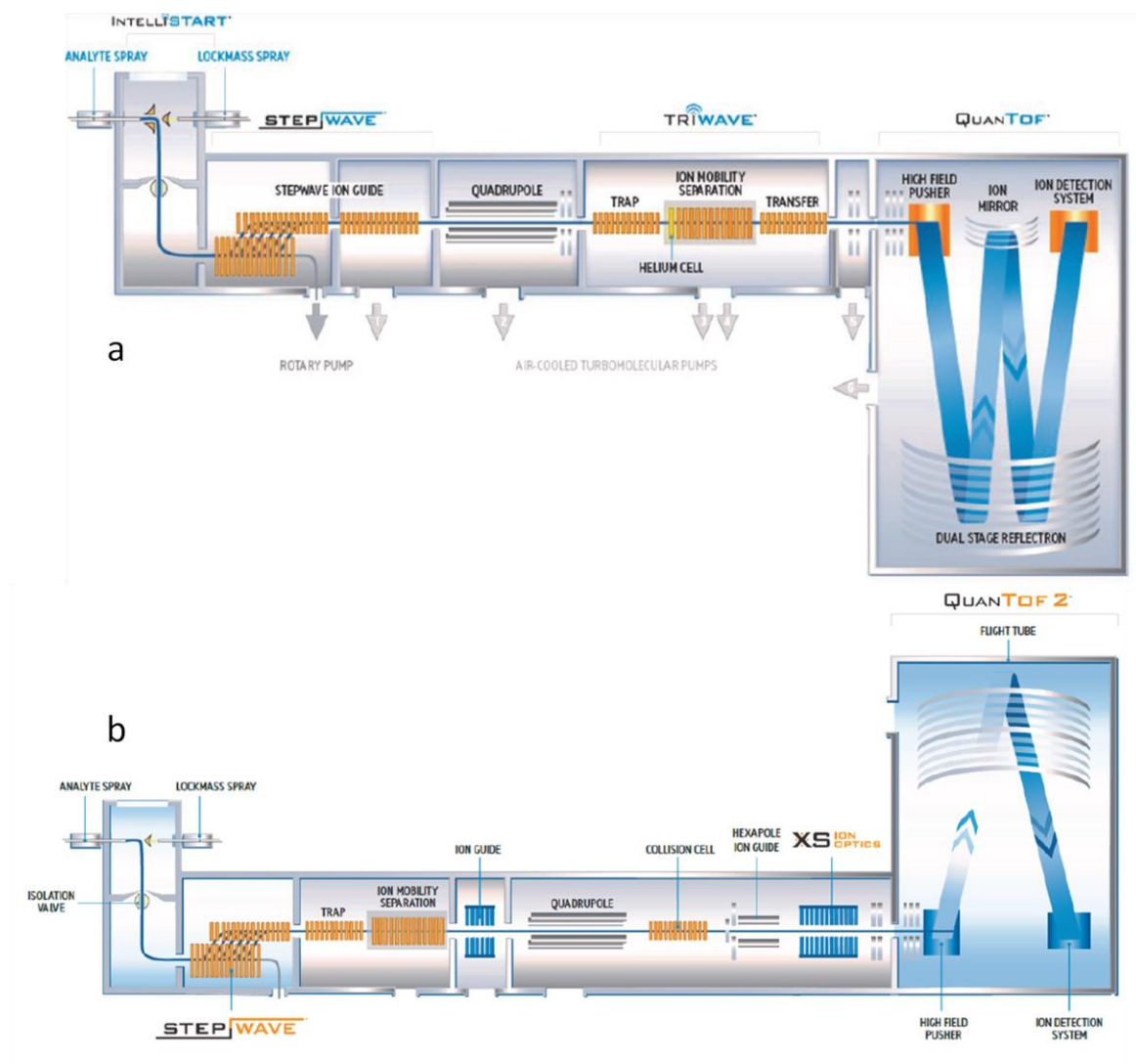


Figure 5 – Schematic of the Synapt G2-Si (a) and Vion (b) TWIMS mass spectrometers commercialized by Waters.

The resolving power of TWIMS (~40) is worse when compared to typical DTIMS analysers, however its duty cycle and sensitivity are much greater. Another convenient facet of the Synapt instrument series is the presence of collision cells before and after the mobility cell (denoted TRAP and TRANSFER, respectively), enabling mobility separation of either precursor or product ions.

2.3 (High-)Field asymmetric (FA)IMS

FAIMS or differential ion mobility spectrometry, unlike DTIMS and TWIMS, operates at high field (high E/N , $>30 \text{ Td}^{[34]}$) and therefore CCS cannot be derived from the mobility data. As a result, FAIMS is always used as a gas-phase separation technique at the front-end of the mass spectrometer. FAIMS achieves ion separation based on the differential mobility operating in low field and high field conditions. Ions are carried by a drift gas between two parallel plates; one plate is held at ground whilst an asymmetric waveform is applied to the other. This waveform consists of a high potential (high-field) for a short time period followed by application of a low potential of opposing

polarity for a longer period of time. These time periods and potentials are designed so that $(V_h t_h) + (V_l t_l) = 0$ where V_h and V_l are the voltages applied during the high-field and low-field period respectively, and t_h and t_l are the time periods for the high-field and low-field portions of the waveform respectively. This ensures that mobility separation is resulting solely from the difference in mobility at high- and low-fields.^[35] A compensation voltage (CV) is applied allowing transmission of ions that have been displaced through their differential mobility by a specific amount. As a result, FAIMS can operate as an ion mobility filter akin to quadrupoles acting as m/z filters. In this mode the sensitivity is much greater (higher duty cycle) as it can separate ion beams rather than ion packets. Conversely the compensation voltage can be scanned to allow detection of all ions, although this reduces the sensitivity. The other major advantage of this technique is that it currently has one of the highest reported resolving powers that has been reported for an IMS device (>500),^[36] with resolving powers routinely reported to be ~ 100 . Like TWIMS, the drift gas employed in FAIMS separation is typically nitrogen to prevent electrical breakdown,^[37] however helium or hydrogen can be doped into nitrogen, which was reported to improve the resolving power of the instrument.^[36] Alternatively, chips with narrower channels could allow the use of helium.^[38]

2.4 Trapped IMS (TIMS)

The final IMS technique that shall be discussed is the recently developed trapped IMS (TIMS). Ions are pulsed (with a deflection plate) into an ion funnel, where they are radially focused before entering the IMS region (Figure 6).^[39] Ions enter the TIMS region, which consists of a series of airtight plates segmented into quadrants, with a flow of drift gas. An axial electric field opposing the drift gas flow is applied trapping ions at different regions within the TIMS cell depending on the ions Ω/z . Simultaneously, an RF potential is applied to the quadrants, creating a quadrupole field perpendicular to the electric field. This quadrupole field restricts ions to the centre of the cell. The axial electric field is then steadily reduced eluting species from high to low Ω/z ^[39a] meaning TIMS can be used as an IMS filter, like FAIMS. Since TIMS operates at the low-field limit, CCS values can be extracted from the data (*via* calibration to known CCS standards), similar to TWIMS. Considering its only recent development and commercialization there are still very few publications in the area, especially those related to carbohydrate separation. However, TIMS has been reported to offer high resolving powers (>250), surpassing that of the other IMS devices (DT- and TW-IMS).^[40]

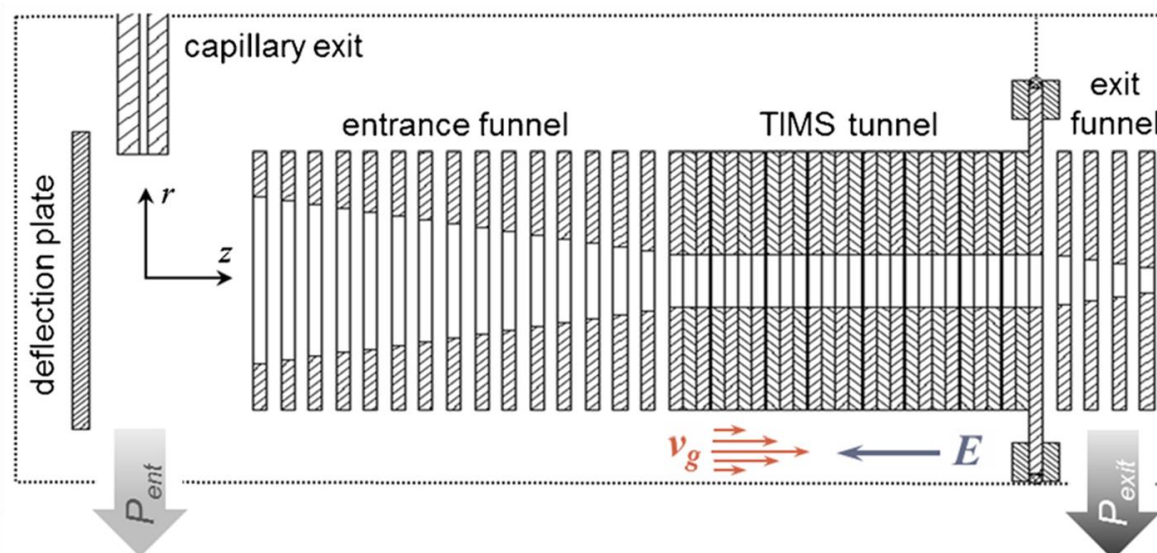


Figure 6 – Scheme of the TIMS cell developed by Bruker that can be coupled onto the front-end of most instrumentation.

3. Computational methodologies

IM measurements alone do not directly reveal any structural insights beyond the relative size of measured ions and their conformational dynamics under the defined set of experimental conditions used. Instead, the CCS of computationally derived structures is compared to the experimental values to elucidate structural information. Theoretical CCS of an ensemble of structures that fit within the CCS distribution may indeed represent the gas-phase structure of the given analyte, assuming the energy of the structure is not unrealistically high. Normally, a set of structures is extracted from the conformational search, with particular focus on the more likely, low energy species. The information gathered from IM-MSⁿ experiments combined with computational techniques such as *ab initio*, semi-empirical or molecular dynamics simulations can offer powerful insights into the physical features of the molecules, including their connectivity, relative size and conformational dynamics.^[41]

Despite significant progress being made in the field, generation of feasible models is not a trivial task, and goes beyond the scope of this review. For large macromolecular species, such as proteins, the initial modelled structures are normally taken from X-ray crystallographic databases or NMR atomic coordinates, as structure elucidation from primary sequence information alone is extremely difficult.

Modelling of smaller molecules, such as glycans, is achievable although computationally challenging even for mono- and di-saccharides. As expected, the main difficulty comes from the high diversity and flexibility of these species.

The CCS values for the generated models are normally based on one of the following theoretical methods; Projection Approximation (PA),^[42] Exact Hard Sphere Scattering (EHSS),^[45] Trajectory Method (TM)^[43] or the Projection Superposition Approximation (PSA)^[44] (Figure 7).^[24b] The choice of algorithm largely depends upon the physical properties and size of the analyte but also the computational resources available. The PA was the first theoretical method used to calculate CCS from atomic coordinates. Simplistically, the molecule is held in a fixed position and the atomic ‘shadow’ it casts onto a 2-dimensional plane is probed with a buffer gas; if the buffer gas was to collide with the ‘shadow’ the collision is counted as a hit and is measured. The molecule is randomly rotated around its axis, which is repeated multiple times to generate a rotationally averaged shadow or projection of the shape of the molecule, and is stopped once defined convergence criteria are met.^[42] The PA method is computationally least demanding, and offers the simplest and quickest approximation of the CCS, however, it does not account for the multiple scattering events that might occur during the collision and ignores all long-range interactions with the buffer gas. Slightly more sophisticated of the theoretical methods is the EHSS which treats molecules and the drift gas as rigid hard spheres, with pre-determined radii. Unlike PA, this method takes into account the multiple scattering events, although still does not take into account the interaction potentials of the system with the buffer gas. As a result EHSS remains computationally inexpensive, whilst offering better approximation.^[45] The most commonly used method to study small molecules is the TM, which accounts for the long-range interactions during the collisions of the molecule with the buffer gas. The TM is typically the most reliable algorithm to theoretically derive CCS, however, it is also the most computationally demanding.^[43] The TM has been parameterized and evaluated for helium and nitrogen, which are typically used in the IMS experiments; the parameters defined in the TM as implemented by Mesleh (for helium)^[43] and Campuzano (for nitrogen)^[46] are limited to several atomic types, meaning only certain classes of molecule can be accurately examined. The typical approach to overcome this limitation has been to define the parameters for unknown atoms yourself, or replace them with similarly sized atoms (compromising the accuracy of the calculated CCS). Finally the PSA, which was developed by Bleiholder *et al.*, includes atomic potentials and a shape factor within the more simplistic and computationally efficient PA algorithm.^[44] This method outperforms the EHSS and agrees relatively well with the TM, however is computationally 100-1000 times quicker making it an attractive alternative for larger systems such as glycoproteins or polysaccharides.^[44a]

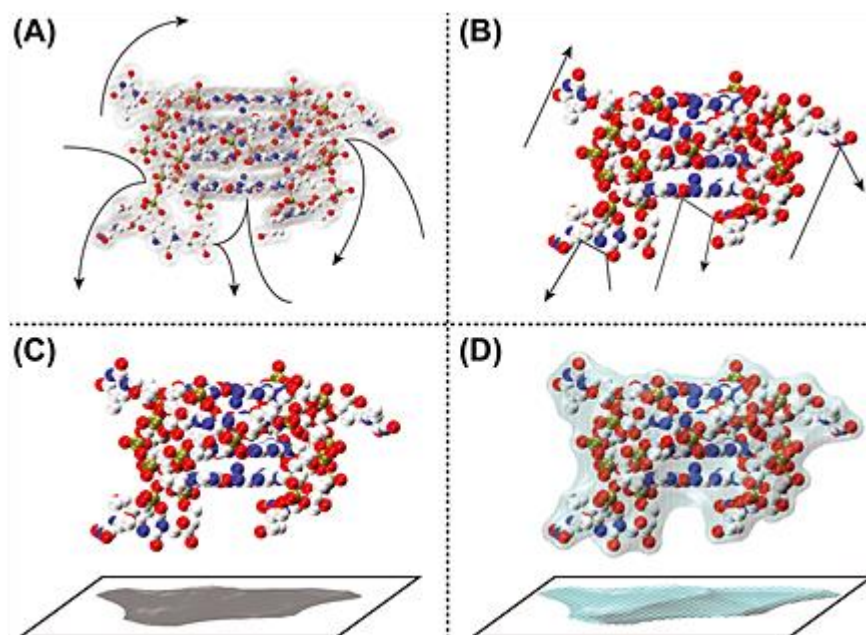


Figure 7 – Schematic of how the different models calculate CCS. The trajectory model, where the long range interactions between the drift gas and analyte ion are taken into account when determining the gas deflection trajectories and scattering angles (a). Exact hard sphere scattering model, where atoms within the analyte are considered as hard spheres and scattering angles between the drift gas and analyte are calculated (b). Projection approximation where the atoms are considered as hard spheres and the projected ‘shadow’ of all possible analyte orientations is calculated (c). Finally, the projection superimposition approximation which is similar to the PA, except atoms are considered as ‘soft’ spheres, accounting for the collective size effect by superimposition of all atomic contributions. The resulting value is scaled by a shape factor to account for molecular cavities (d). Figure taken from D’Atri *et al.*^[24b]

4. Application of IMS to analysis of carbohydrates and their conjugates

Broadly speaking, IMS can be applied for two functions: separation of ions and the measurement of arrival time distributions (ATDs) and calculation of CCS (structural applications) in the hope of defining precise gas-phase structural information from computational studies. However, there is currently no specific set of IMS conditions that have been demonstrated to be able to separate all carbohydrates or computational methods to generate theoretical structures. As a result, IMS separation of derivatized^[47] and underivatized carbohydrates have been performed with a plethora of different adducts in both positive and negative ion mode.^[48] Separation has been achieved with a variety of drift gases,^[48b, 49] at different temperatures and pressures, on different mobility analysers.^[49a, 50] Samples have also been ionized by different techniques, predominantly MALDI^[48d] and ESI,^[50b] with differing sample preparations, all of which will affect the measured ATDs. Improved separation has been reported in numerous ways for each IMS device. Changing the drift gas to larger, more polarizable molecules such as carbon dioxide has resulted in the ability to baseline separate isomers, which were otherwise unresolved.^[48b, 49a, 51] Lalli *et al.* reported for other small molecule isomers that increasing the pressure also improves the resolving power (rather than peak-to-peak

distance), although massively impacts on the ion transmission (*i.e.* sensitivity).^[49a] Raising the temperature of the drift gas is also likely to reduce the resolving power based on theoretical calculations, although no systematic study has been attempted for carbohydrates as far as the authors are aware.^[52] Additionally it is (currently) impossible to discern any structural information from computational calculations using polarizable gases as no suitable algorithms have been formulated.^[49a] Alternatively, changing the ions adduct type can result in differential mobilities.^[53] These factors have allowed the separation α/β -anomers of 1-Me-Glc, 1-Me-Gal and 1-Me-Man.^[48b] Recently, Hofmann and co-workers reported the greatest separation of anomers and regioisomers as deprotonated adducts compared to protonated, sodiated and chloride adducted.^[48a] Separation as metal adducts may be more challenging as the structures ‘deform’ in the gas-phase in order to solvate the charge of the adduct,^[54] potentially masking isomeric differences in certain cases. Computational studies can also be more challenging when using more ‘exotic’ adducts (such as heavy metals) for reasons similar to those occurring with different drift gases. Improvements in IMS separation achieved by changing the drift gas or adduct are difficult to predict as they depend on the chemical properties of the analyte.^[55] In fact the choice of adduct is normally dictated either by the analyte, (acidic carbohydrates normally deprotonate readily^[56] whereas neutral carbohydrates tend to sodiate^[50b]) or its ability to generate rich, structurally informative and diagnostic product ions upon activation of the complex.^[57] Applications of IMS to facilitate separation and structural characterization of glycans will be discussed in detail for the different types of glycans and glycoconjugates.

4.1 Free-glycans

4.1.1 Mono- to hexa-saccharides

4.1.1.1 Separation

Initial application of IMS to carbohydrates focused on whether or not IMS had sufficient resolution to distinguish simple (mainly mono-, di- and tri-saccharides) isomeric carbohydrates. Liu *et al.* (1997) first demonstrated that ability of DTIMS_{N₂} to distinguish a small sub-set of isomeric precursor and product ions derived from raffinose and melezitose and α -cyclodextrin, β -cyclodextrin and γ -cyclodextrin.^[58] Using their home-built DTIMS_{N₂} device, the Hill group then demonstrated separation of 21 different carbohydrate standards, including a significant number of isomers.^[59] The same group later reported the ability to separate a substantial number of all known (sodiated) 1-methyl hexoses and selected 1-methyl N-acetylhexosamines (GlcNAc and GalNAc) monosaccharides as both their α/β - anomers using both DTIMS and TWIMS. For the most part α - and β - 1-methyl anomers of each hexose could be discriminated by both DTIMS and TWIMS other than gulose and certain diastereoisomers could also not be separated such as 1-Me- β -allose and 1-Me- β -glucose.^[60] As might be expected the DTIMS displayed a higher resolving power. Recently, the Pagel and Seeberger groups demonstrated the ability of IM-MS to separate and quantify anomers and regioisomers of

chemically synthesized tri-/penta-saccharide precursor and product ions down to concentrations as low as 0.1% of the dominant anomer glycans (Figure 8).^[48a]

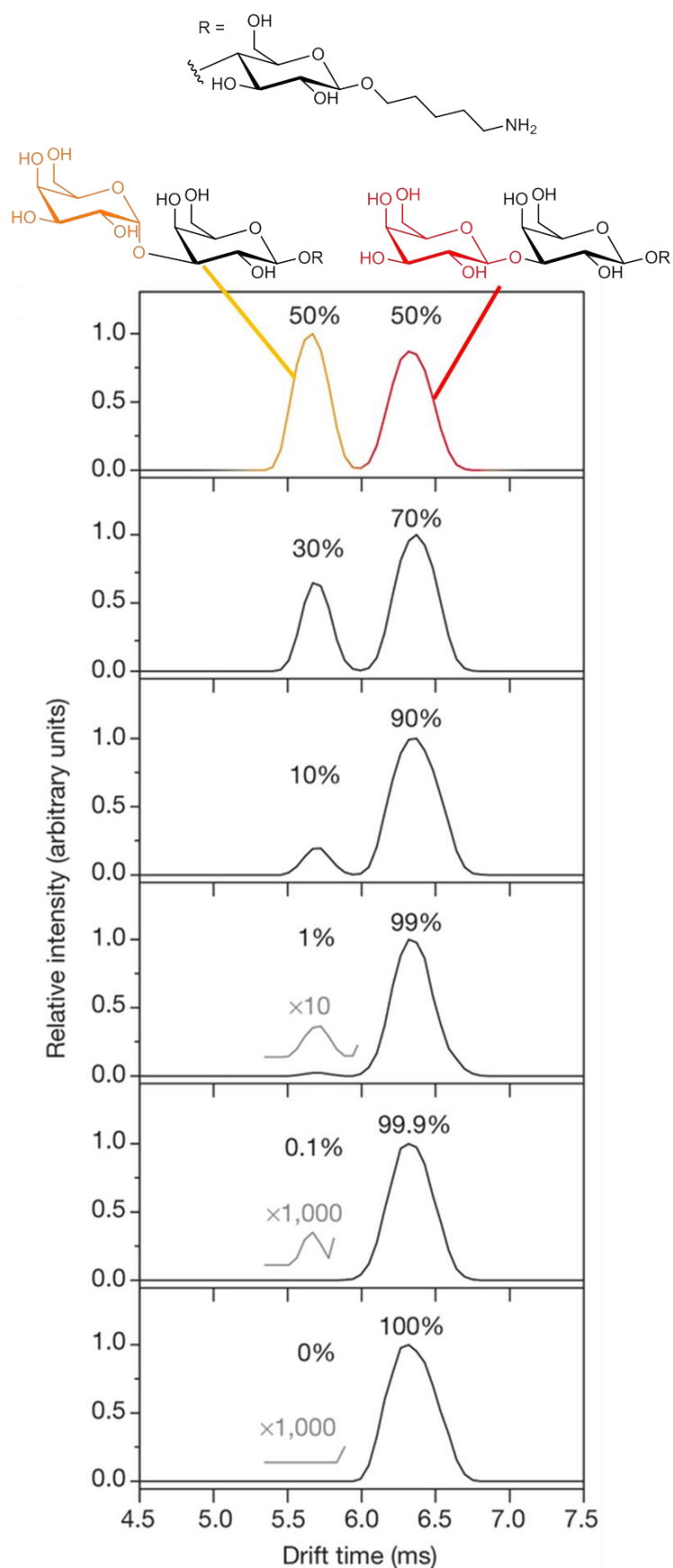


Figure 8 – ATDs associated with a mixture of two anomeric carbohydrates at ratios defined on each distribution. Figure amended from Hofmann 2015.^[48a]

FAIMS has also been demonstrated to be capable of resolving anomeric, linkage and positional isomers of a series of chemically synthesized glycosides as a number of different adducts in both positive and negative ion mode.^[48c] Mobility separated precursors are routinely dissociated by collision-induced dissociation (CID) to generate structural information.^[57a, 61]

Even with application of IMS, baseline separation of isomeric precursors is often not fully achieved. Chimeric ATDs resulting from isomeric precursors can be extracted if a post-mobility generated product ion is specific to a given isomer (sometimes termed extracted fragment ion drift time distributions or XFIDTDs).^[53, 61e] Post-mobility CID enabled elucidation of isomeric glycosides within the glycourinome of a patient suffering from symptoms assigned to a congenital disorder of glycosylation.^[62] This structural variance could have been overlooked by MS alone. Alternatively, the mobility of product ions generated from CID of isomeric precursors may be separable.^[53, 61c, 63] For example Li *et al.* (2013) measured the ATDs of a series of dihexose anomers, regioisomers and diastereoisomers by TWIMS. Despite the fact that certain precursors co-eluted in mobility, their respective monosaccharide-glycolaldehyde cross-ring product ions did not.^[63] To circumvent the possibility of the quadrupole co-selecting isomeric/baric precursors for fragmentation, which would result in unwanted chimeric product ion spectra, the same group incorporated a dual gated DTIMS device in place of the source region of a Synapt HDMS (commercial TWIMS instrument). This allows mobility selection of precursors prior to mass selection, CID and subsequent mobility separation of the generated products ions (tandem IMS). This mobility selection was achieved using a dual gate drift tube where one gate allows ions to enter the drift cell and the second allows ions to leave the cell. The second gate is opened at a defined time after the first allowing only ions of that specific mobility to be transferred into the ion trap. However, this gating of ions dramatically reduces the duty cycle and by virtue the sensitivity (Figure 9).^[64] Further improvements in separation has been achieved by combining energy-resolved (ER)-MS with IMS, since diastereomeric glycans may dissociate at different energies.^[57b, 65]

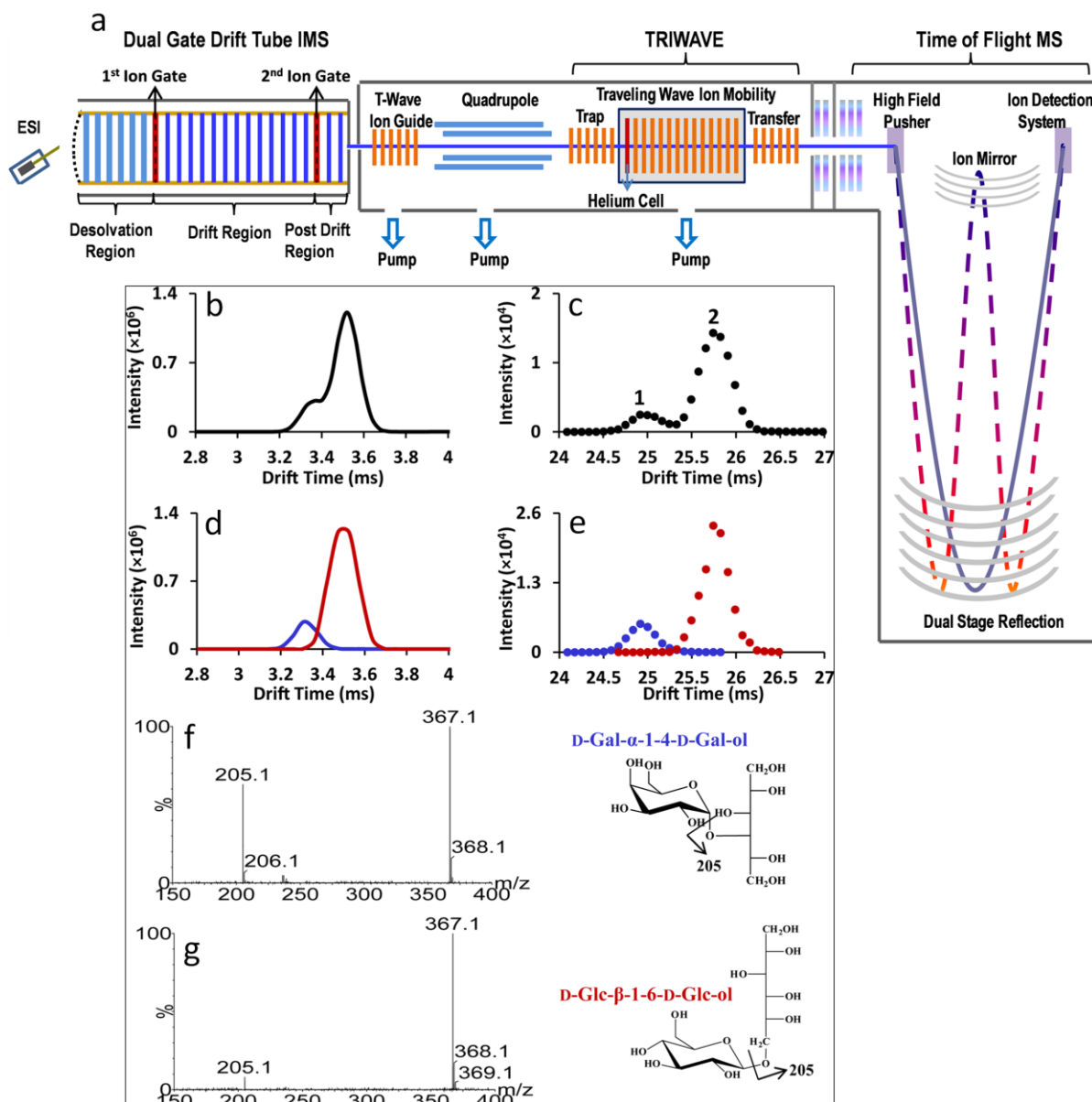


Figure 9 – The tandem IMS instrument developed by the Hill group, formed from connecting a dual gated DTIMS onto the source of a TWIMS mass spectrometer (a). Inset are the DT (b) and TW (c) ATDs associated with a mixture of Gal α -1-4Gal-ol and Glc β 1-6Glc-ol. ATDs of the pure compounds measured by the DT (d) and TW (e) analyser confirming that the mixture is separable. (f) and (g) are the tandem mass spectra associated with the mobility selected peaks labelled 1 and 2, respectively, within panel (c). Figure amended from Li *et al.*^[64]

From initial work on simple chemically synthesized standards, isomeric disaccharide-alditols, chemically released from the *O*-linked glycoprotein bovine submaxillary mucin (BSM) by alkaline borohydride treatment, have been separated by IMS, which could not be achieved by HPLC alone.^[61b, 66] Unlike reducing carbohydrates these alditols possess less conformational variability (*i.e.* narrow ATDs consisting of a single peak), which is likely as a result of the inability of non-reducing glycans to mutarotate between isomeric structures.^[61a, 61b] MS^{*n*} (multistage tandem mass spectrometry where *n* denotes the number of activation events) of mobility selected HPLC fractionated BSM carbohydrate-alditols, yielded further diagnostic structural information.^[61a, 61b] Anionic carbohydrate alditols were

also separable by TWIMS with greater sensitivity, albeit at the loss of some IMS resolving power.^[61c] Post-mobility CID generated diagnostic tandem mass spectra from isomeric precursors. Additionally, CID generated BSM carbohydrate-alditol product ions could be separated with and without mass isolation. The mobility separated product ions could also be made to dissociate again producing further structural information.^[61c]

As the reader may have noticed, the vast majority of these strategies exploit CID to fragment precursors, principally because it is the simplest method to achieve dissociation. However, other informative dissociation methods have also been employed. Lee *et al.* performed vacuum ultraviolet photodissociation (VUVPD) (157 nm) on several mobility-separated isomeric disaccharides, which produce similar CID tandem mass spectra.^[57a] VUVPD generated significantly more cross-ring fragments than CID and their presence was found to be highly dependent on the regiochemistry and anomeric configuration of specific precursors. This enabled discrimination and identification of all isomers.^[57a] Finally Hernandez *et al.* mobility selected lithiated, sodiated and potassiated α/β -1-methyl anomers of Glc and Man by FAIMS.^[61d] Mobility selected ions were accumulated in a quadrupole ion trap before being irradiated with various wavelengths of infrared radiation. Ions absorb IR radiation and dissociate when their structures possess resonant vibrational modes. Therefore plotting the fragmentation yield against the wavelength of light, normalized to the respective laser power at each frequency, generates an action IR spectrum. From this action IR spectrum, the gas-phase structures of each of these complexes were determined in the presence of a single water molecule (Figure 10).^[61d] The combination of IMS, IR and MS is an extremely powerful technique. IMS can separate isomers, which would be indistinguishable by MS alone and can provide structural information on the intact complex; MS rapidly separates species by their m/z and can generate connectivity information through tandem mass spectrometry; IR provides specific atomistic information often lacking from IM-MS approaches alone.

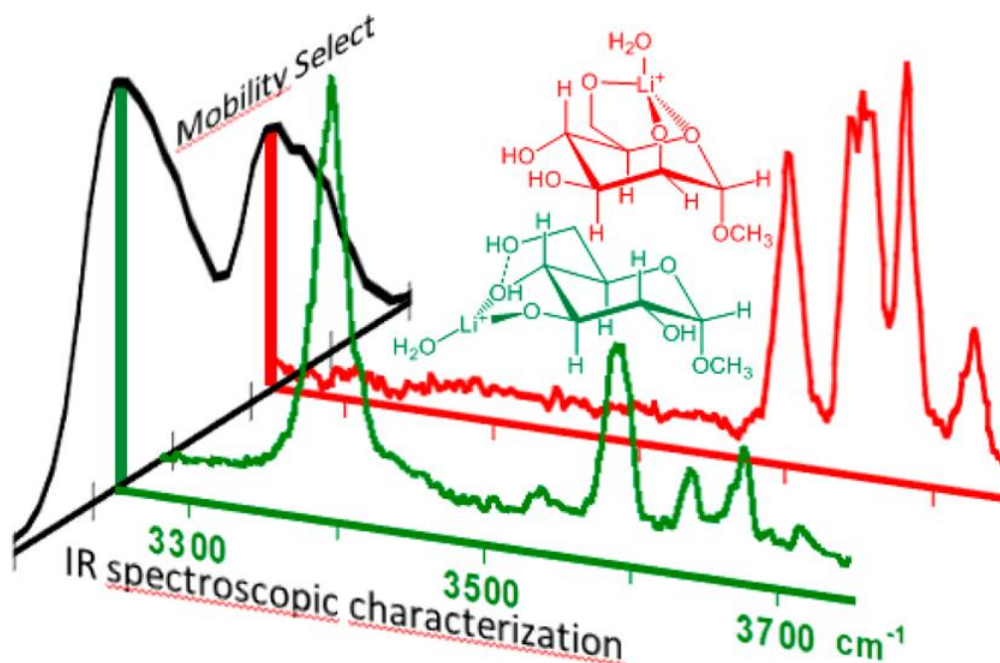


Figure 10 – $[\text{Mea1-Glc}+\text{H}_2\text{O}+\text{Li}]^+$ (green) and $[\text{Mea1-Man}+\text{H}_2\text{O}+\text{Li}]^+$ (red) complexes, mobility separated by FAIMS (black trace). Species can then be mobility selected prior to variable wavelength action IR MS. MD calculations of theoretical structures that would give rise to these traces suggest that the coordination site of Li^+ differs between Mea1-Glc and Mea1-Man. Figure taken from Hernandez.^[61d]

IMS provides benefits, other than just separating isomeric ions: it can be used to help ‘filter’ carbohydrate specific data from contaminating ions or other molecular species.^[62] This can be easily visualized on a plot of m/z vs. drift time/CCS as certain classes of ions tend to run in similar regions on this plot. This enables simultaneous analysis of glycans and glycopeptides within the same experiment, circumventing laborious sample purification and separation steps and potentially facilitating future high-throughput glycoproteomics.^[67] Carbohydrates can also be chemically derivatized to alter their m/z and CCS, so they appear in an uncongested region within this 2D plot.^[47]

4.1.1.2 Structural

As mentioned within the introduction of this section, IMS can provide structural information, facilitated by computational calculations. This was first reported by the Bowers group, who measured the $^{\text{DT}}\text{CCS}_{\text{He}}$ of a series of sodiated tetrahexoses and hexoses. The lowest energy conformations of these structures were computed and their theoretical CCS_{He} determined by projection approximation. For each carbohydrate, the lowest energy structures had the carbohydrate chain wrapping around the sodium ion, forming 4 coordinate bonds to 4 monosaccharide units, solvating the charge. These theoretical CCS_{He} matched well with experimental values suggesting that these were indeed reasonable representations of the measured gas-phase structures.(Figure 11)^[54]

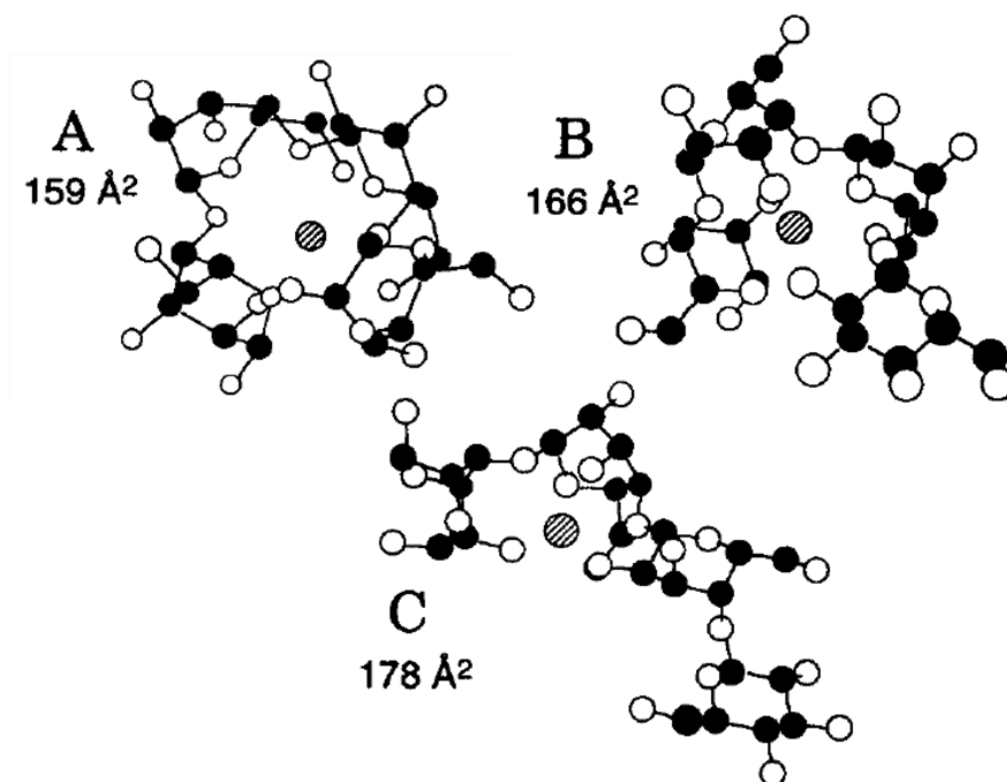
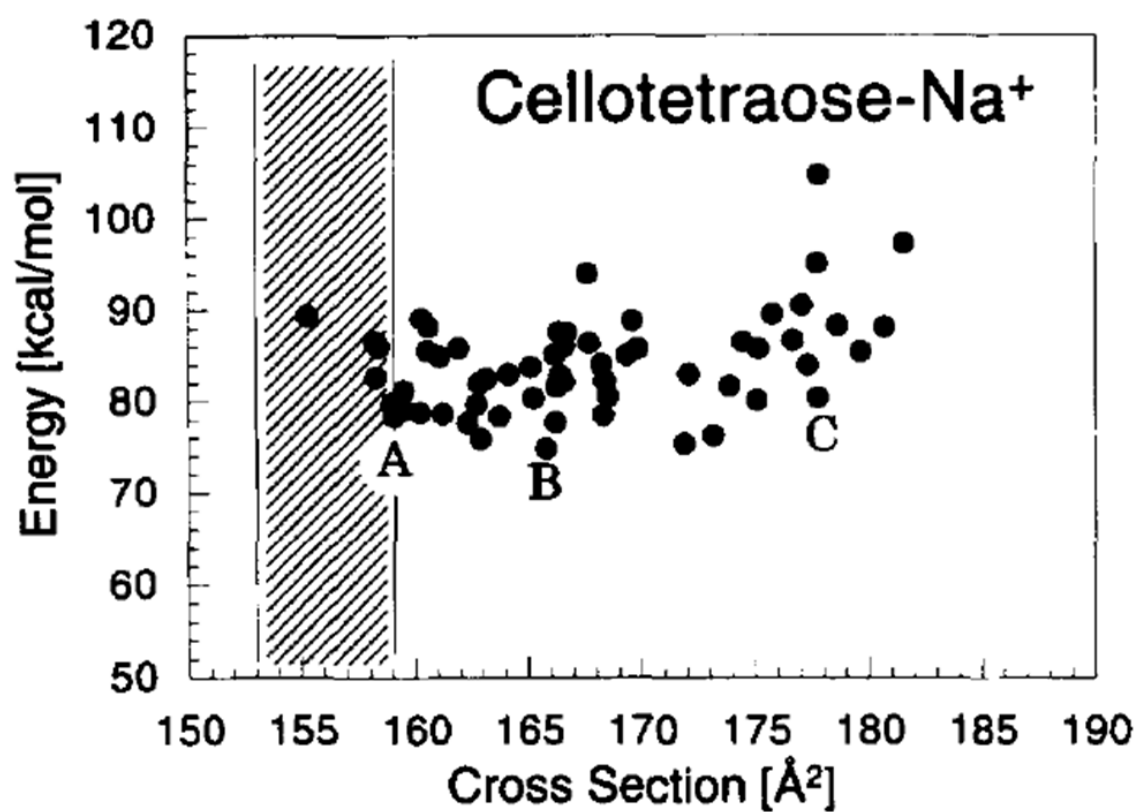


Figure 11 – Scatter plot representing the energy and cross section of MD generated sodiated callotetraose structures. The shaded area highlights the experimentally measured ^{DT}CCS_{He⁺}. Structures A, B and C are representative structures within areas of the plot, Figure amended from Lee *et al.*^[54]

Leavel *et al.* showed by DTIMS that for a $[\text{Zn}+\text{diethylenetriamine}+\text{hexose}+\text{Cl}]^+$ complex the stereochemistry of the hexose also influences the metal binding site.^[55a] A significant number of recent papers have employed TWIMS to measure the CCS of carbohydrate ions. As mentioned in section 2.2 CCS measurements can only be made on TWIMS instruments by comparing it to a calibration curve of standards whose CCS areas had been previously measured on a DTIMS instrument. However, care must be taken when choosing appropriate standards as it has been reported that calibrating glycan CCS to a peptide calibrant introduces a small but systematic error of ~1%, presumably due to differences in the interaction potential between peptides and carbohydrates with the drift gas.^[31] Similarly, charge state mismatching introduces large errors (~3.5%), *i.e.* the error associated with normalising both mass and charge states within the measured data. Mismatching both charge state and the molecular class increases this error further (~4.7%).^[31a] Similarly, large errors are associated with trying to convert $^{\text{DT}}\text{CCS}_{\text{He}}$ to $^{\text{TW}}\text{CCS}_{\text{N}_2}$.^[31b] Fenn *et al.* measured the $^{\text{DT}}\text{CCS}_{\text{He}}$ values for over 300 carbohydrates including isomers,^[53] and Hofmann *et al.* measured the $^{\text{DT}}\text{CCS}_{\text{He}}$ and $^{\text{DT}}\text{CCS}_{\text{N}_2}$ of a dextran polymer^[31b], both of which can be used to calibrate TWIMS analysed carbohydrates. Pagel *et al.* also determined the $^{\text{DT}}\text{CCS}_{\text{He}}$ and $^{\text{DT}}\text{CCS}_{\text{N}_2}$ of N-glycan precursors and product ions. Glycans were released with hydrazine from RNase B, fetuin, asialofetuin, ovalbumin, porcine thyroglobin and desialylated porcine thyroglobin, which could be beneficial for calibration of TWIMS for N-glycan analysis as discussed within the next section.^[68]

Huang *et al.* measured the $^{\text{TW}}\text{CCS}_{\text{N}_2}$ of various di-, tri-, penta- and hexa-saccharides as their group I metal adducts (Li^+ , Na^+ , K^+ , Rb^+ , Cs^+) and noted that the $^{\text{TW}}\text{CCS}_{\text{N}_2}$ generally increased as the metal ion radius increased, although in an isomer specific manner, which likely reflects the differences in coordination site.^[53, 55b] The same group also demonstrated IMS separation of isomeric lacto-N-fucopentose I and lacto-N-fucopentose V, and lacto-N-difucohexaose I and lacto-N-difucohexaose II as Ca and Ba radical cation adducts,^[69] which have been reported to be difficult to separate by both IMS and MS^2 , including as divalent Ca^{2+} and Ba^{2+} adducts.^[53, 55b, 69-70] An additional benefit was that these radical cations generated much more diagnostic CID product ions compared to their divalent equivalent.^[69]

Structure determination is not only limited to precursor ions, but also product ions. This is exemplified by the work of Both *et al.* who measured the TWIMS arrival time distributions of a series of isomeric hexose and N-acetylhexosamine glycosidic product ions (B-, C-, Z- and Y-) derived from CID of either glycopeptides or reducing carbohydrates.^[50b] Crucially, the ATDs of these product ions were independent of the initial precursor, therefore they were able to ‘sequence’ the stereochemistry of the monomeric product ions (Figure 12). Incorporation of multistage tandem mass spectrometry prior to IMS enabled complete elucidation of the monosaccharide building blocks within Man3 glycan and helped reveal promiscuity of the glycosyltransferase pp-GanT2.^[50b]

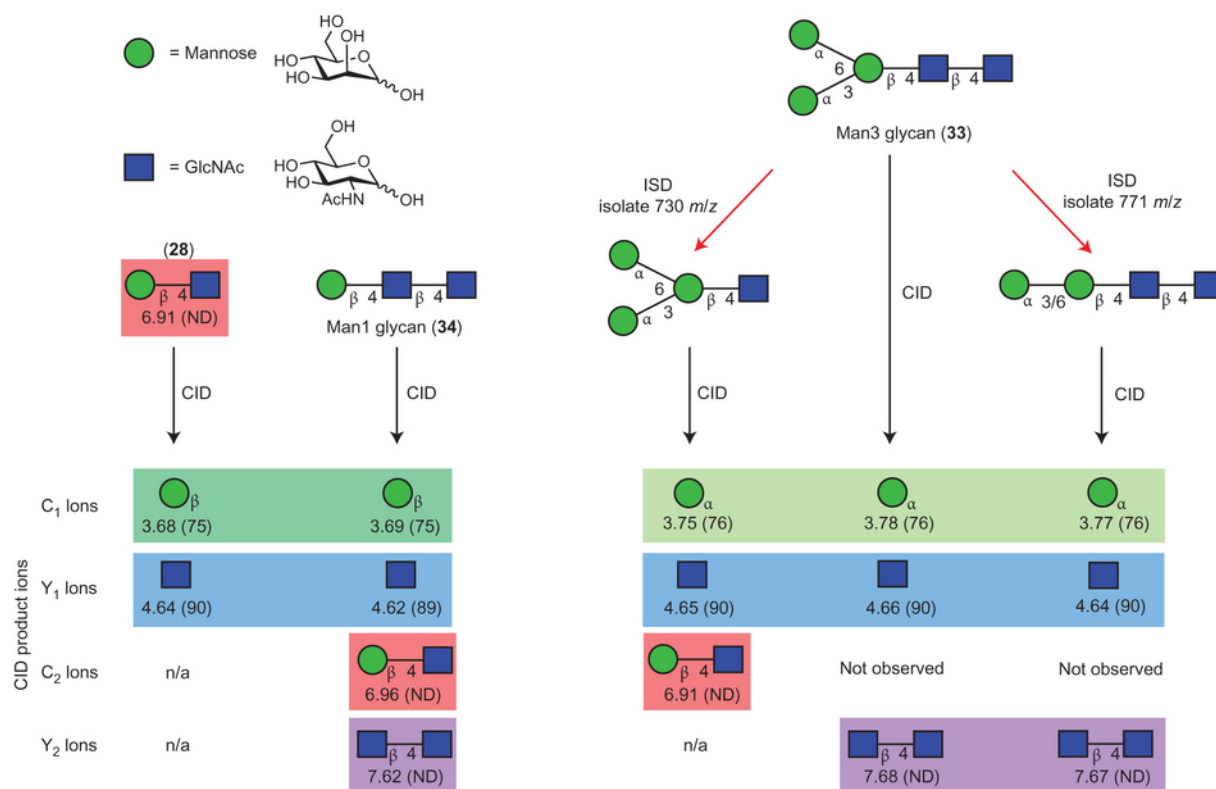


Figure 12 - Drift times (in ms) and CCS values (in Å²) in parentheses for selected [M+Na]⁺ product ions are presented below the structures. Terminal monosaccharide and disaccharide product ions could be characterized after a single round of CID. Internal residues were characterized following in-source dissociation (ISD), quadrupole mass isolation of the desired ISD product ion, and subsequent CID before IM-MS. C₁ ions can be grouped either as β-Man (shaded dark green) or α-Man (shaded light green). All identified Y₁, C₂ and Y₂ product ions could be characterized as GlcNAc (shaded blue), Manβ1-4GlcNAc (shaded red) and GlcNAcβ1-4GlcNAc (shaded purple), respectively. For cases where two conformations (potential sites of sodiation) were observed, the drift time and CCS measurements are reported for the predominant species. CCS values for the disaccharides were not determined (ND) as the drift time values lie outside this calibration range. n/a, not applicable. Figure taken from Both *et al.*^[50b]

Recently, the Clemmer group measured the ^{DT}CCS_{He} of lithiated precursor and CID generated product ions on a home-built tandem DTIMS instrument. Even though specific precursors had identical CCS (for example sucrose and leucrose) their product ion CCSs differed (although these product ions may have consisted of multiple different species due to the potential existence of isomeric product ions).^[71]

4.1.2 N-glycans

Analysis of structurally larger and more complicated N-glycans largely built upon the initial studies on smaller glycan standards.^[67-68] Fenn *et al.* and Harvey *et al.* demonstrated the benefit of the additional gas-phase separation afforded by IMS to help characterize N-glycan isomers released from the glycoproteins RNase B, gp120 and fetuin, either enzymatically or chemically.^[48d, 56, 67] Zhu and co-workers were able to separate four isomers of permethylated Man7 glycan released from RNase B by DTIMS and post-mobility CID. Deconvolution of the ATDs into the four isomers was facilitated by extracting the ATDs associated to isomer specific fragment ions.^[61e] Like for small molecules, N-glycans can also be readily separated from contaminating species.^[56, 72] For example N-glycans have been profiled directly following PNGase F release in the presence of salt and detergents. This

minimizes sample loss from purification strategies or liquid handling steps and additionally reduces the experiment time.^[56, 72b] Glycosides can also be readily deconvoluted into classes from the m/z vs. drift time 2D plots (Figure 13), for example singly, doubly and triply charged glycans and glycopeptides. These species would be unidentifiable based on the signal-to-noise of the MS acquisition alone.^[56] Glycans and glycopeptides could also be fractionated by LC prior to IMS to improve separation and the detection capacity.^[72a]

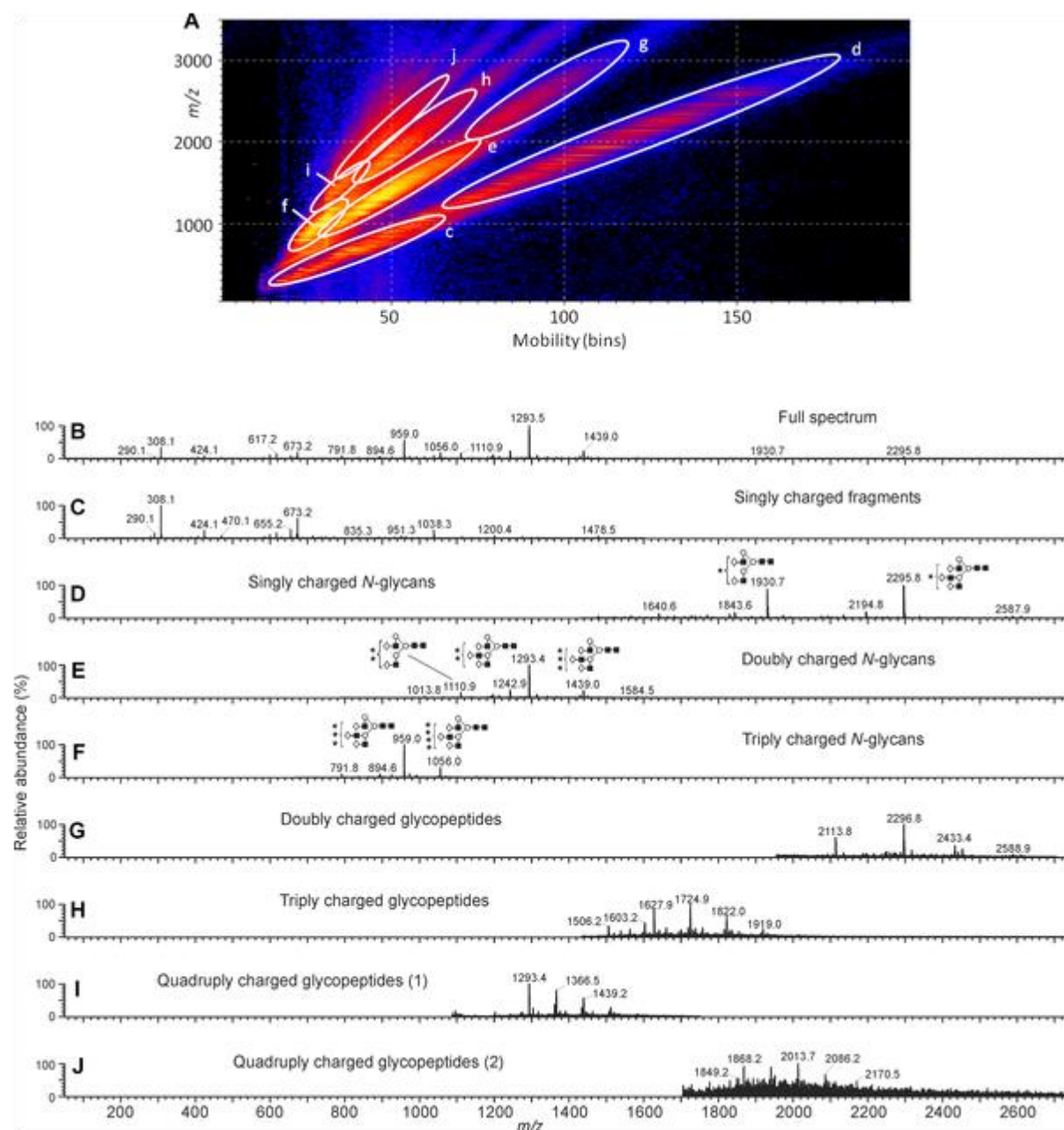


Figure 13 – 2D plot of the drift time against m/z (log intensity scale) of the negative charged ions generated from ESI of N-glycosides released from bovine fetuin by hydrazinolysis (a). The total mass spectrum (b) and the extracted mass spectra of the regions (c-j) within the 2D plot. Figure taken from Harvey *et al.* with permission.^[56]

With the additional orthogonal separation afforded by IMS, Isailovic *et al.* demonstrated profiling of total human serum glycans from patients suffering from liver cancer or cirrhosis of the

liver.^[10c] Differences in the ATDs associated to specific permethylated N-glycan precursor ions, likely result from changes in the isomeric distribution of that given ion. In some cases, variance in the ATDs of certain glycan ions between individuals could be separated into healthy and diseased cohorts after principle component analysis (PCA).^[10c] This data suggests that aberrations in isomer distributions may be indicative of disease states. Considering ATDs associated with multiple different isomeric glycans improved the reliability of the PCA to cluster 81 individual samples into specific disease (and apparently healthy) cohorts significantly (Figure 14),^[10d] whereas MS alone did not demonstrate any significant clustering. Using the same strategy, Gaye and co-workers were able to characterize serum N-glycans into cohorts derived from three disease phenotypes namely Barrett's esophagus, high-grade dysplasia and esophageal adenocarcinoma, from normal controls of apparently healthy individuals.^[10e] Interestingly, a significant number of glycans selected for PCA within this experiment were identical to those selected for the liver disease state experiments.^[10e]

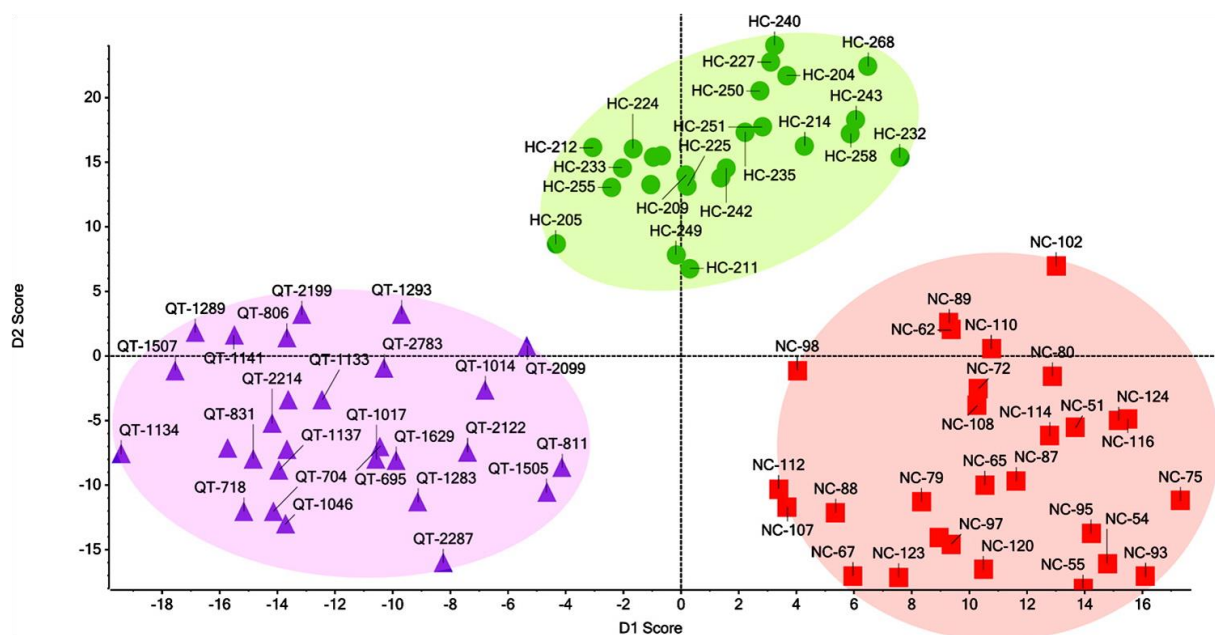


Figure 14 – Supervised PCA of drift time profiles of 10 unique m/z glycan ions namely $\text{Fuc}_1\text{Hex}_5\text{HexNAc}_4^{3+}$, $\text{Neu5Ac}_1\text{Hex}_5\text{HexNAc}_4^{2+}$, $\text{Neu5Ac}_1\text{Hex}_5\text{HexNAc}_4^{3+}$, $\text{Neu5Ac}_1\text{Fuc}_1\text{Hex}_5\text{HexNAc}_4^{3+}$, $\text{Neu5Ac}_2\text{Hex}_5\text{HexNAc}_4^{2+}$, $\text{Neu5Ac}_2\text{Hex}_5\text{HexNAc}_4^{3+}$, $\text{Neu5Ac}_2\text{Hex}_6\text{HexNAc}_5^{3+}$, $\text{Neu5Ac}_3\text{Hex}_6\text{HexNAc}_5^{3+}$, $\text{Neu5Ac}_3\text{Fuc}_1\text{Hex}_6\text{HexNAc}_5^{3+}$, and $\text{Hex}_5\text{HexNAc}_2^{2+}$. Taken from Isailovic *et al.*^[10d]

Given the increased structural complexity associated with N-glycans compared to the smaller glycosides, few papers have been able to deliver structural information from the IMS data. Impressively though, Plasencia *et al.* determined the $^{DT}\text{CCS}_{\text{He}}$ for three species associated with the disodiated permethylated $\text{Hex}_5\text{HexNAc}_4$ glycan released from ovalbumin (Figure 15). They argue that these three species arise from three different isomers of $\text{Hex}_5\text{HexNAc}_4$. CCS calculations (using TM) of the 10 lowest energy structures (after sequential simulated annealing) for each potential isomer were within 2.2% uncertainty of the experimental data, allowing assignment of each species within the ATD for these three isomers.^[73] Williams *et al.* also determined the $^{DT}\text{CCS}_{\text{He}}$ and $^{TW}\text{CCS}_{\text{N}_2}$ for N-

glycans released from RNase B, ovalbumin and porcine thyroglobin with and without desialylation. Using both the EHSS and TM, they determine the CCS of modelled structures, which again match well with the experimental data from both DTIMS and TWIMS analysers.^[33c] Finally, Struwe *et al.* also measured $^{DT}CCS_{He}$, $^{DT}CCS_{N_2}$, $^{TW}CCS_{He}$, and $^{TW}CCS_{N_2}$ for a series of N-glycans as protonated, deprotonated, sodiated, potassiated, chloride and phosphate adducts. Computational simulations suggest that N-glycans wrap around metal adducts to solvate charge, as previously demonstrated by the Bowers group for smaller carbohydrates.^[54] Also for given carbohydrate ions, multiple conformations were observed. These CCS values were stored in a database (Glycomob housed within UniCarbKB) as a Glycomics IMS-MS resource.^[74]

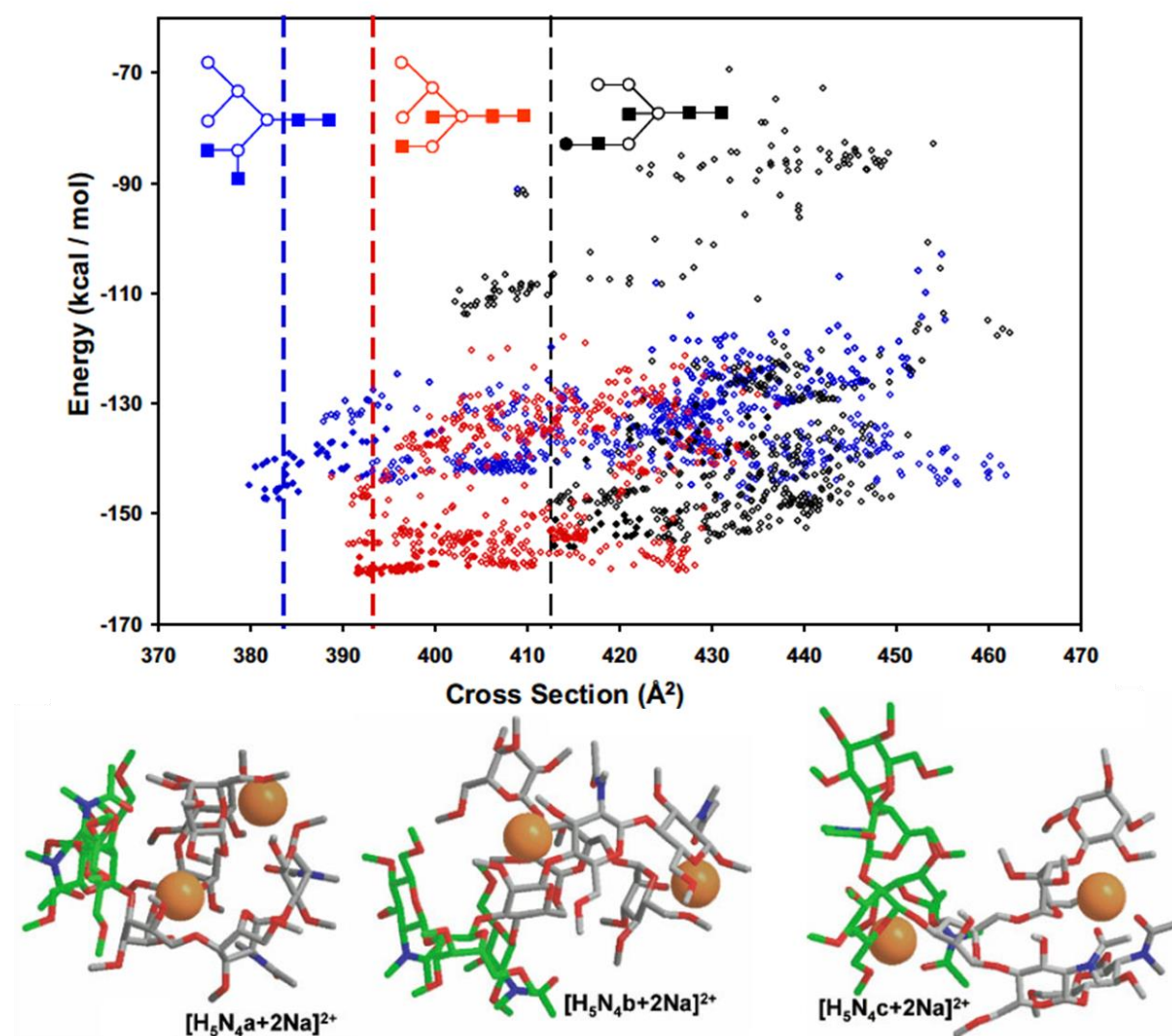


Figure 15 – Calculated cross sections (TM) and energies from MD simulations after simulated annealing for the three $[H_5N_4+2Na]^{2+}$ isomers. Experimental CCS for the three isomers are highlighted by the dashed lines. $[H_5N_4a+2Na]^{2+}$, $[H_5N_4b+2Na]^{2+}$, and $[H_5N_4c+2Na]^{2+}$ are represented by blue, red and black diamonds respectively. Representative low-energy structures for each isomer are shown. Bisecting GlcNAc and the $\alpha 1-3$ mannose arm are shown in green for clarity. Figure amended from Plasencia *et al.*^[73]

4.1.3 Glycosaminoglycans (GAGs) and other Polysaccharides

Glycosaminoglycans (GAGs) are a family of linear anionic polysaccharides found almost ubiquitously on animal cell surfaces and within the extracellular matrices. GAGs have several biological functions such as cell-cell communication, cell adhesion, modulation of axonal and cell growth, proliferation, angiogenesis, anti-coagulation properties, viral invasion, tumor progression and metastasis.^[75] Furthermore, they are known to bind numerous proteins such as chemokines, cytokines, proteases, growth factors, pathogen proteins and adhesion molecules.^[75b, 75c, 76] Consequently, GAGs have an extensive medical potential application (*e.g.* heparin for their anticoagulant properties).^[77] Characterization of this family of polysaccharides is therefore fundamental to understand their structure/function relationship but represents a challenging task due to their inherent sequence, de-acetylation and sulfation heterogeneity.^[11b, 75a, 76a, 78] Consequently, despite the numerous biological activities of GAGs, there are few function specific sequences that have been completely elucidated.^[79] It is also known that modification of the GAG affects its conformation and as a result influences its ability to bind certain proteins such as the fibroblast growth factor-2 or antithrombin that are involved in the blood-coagulation cascade.^[80] IMS offers the ability to monitor potential dynamic conformational changes upon varying the GAG sequence identity. Jin *et al.* performed conformational analysis of fully or partially sulfated heparin-derived di- and tetra-saccharides using a combination of DTIMS-MS and molecular modelling (using PA model) of $^{DT}CCS_{He}$.^[33e] The authors demonstrated that even minor changes to the primary structure of the GAG saccharides, for example changing the position of one sulfate or epimerization of an IdoA residue to GlcA, led to significant changes in conformation. Computational models supported the experimental results, and suggested that sodium adducts localized towards negatively charged groups and thus play an important role in conformation stabilization and the observed oligomerization of disaccharide units.^[33e] The heparin-derived tetrasaccharides wrap around the sodium cations, presumably to solvate charge, forming compact structures.^[54] More recently, Leary *et al.* explored the affect various alkali metal (Na^+ , K^+), alkaline earth metal (Mg^{2+} , Ca^{2+}) and transition metal (Mn^{2+} , Co^{2+} , Fe^{2+} , Ni^{2+}) adducts had upon the conformation of a heparin octasaccharide,^[81] since it has been previously demonstrated that these cations have influenced several protein-heparin interactions.^[82] In all cases, increased affinity resulted in contraction of the heparin structure. However, conformational changes upon successive addition of metal cations depended on the identity of the metal adduct, which probably results from differing atomic radii and valences (Figure 16). In particular, the Ca^{2+} ion induced significant conformational contraction. Importantly, similar conformational changes were observed in solution to the gas phase IM-MS experiments, illustrating the relevance of the IMS techniques to investigate the structure/function relationship of GAGs.^[81]

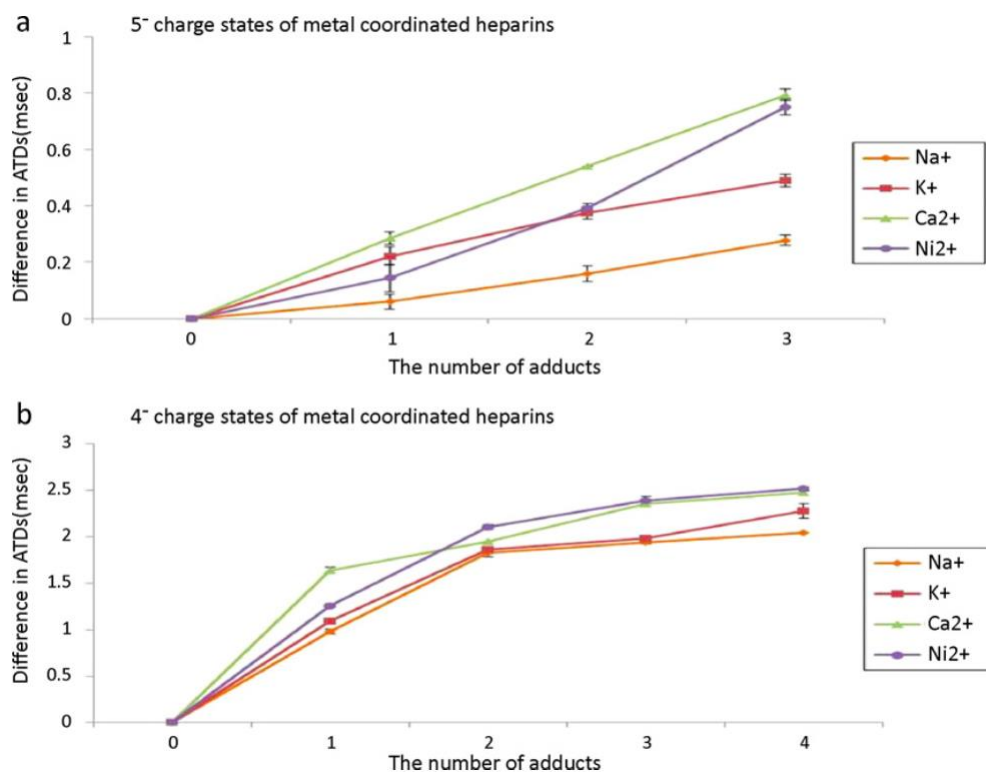


Figure 16 - The difference in arrival time distributions between metal and metal free coordinated heparin octasaccharide, plotted against the number of metal adducts (a) in the 5⁻ charge state and (b) in the 4⁻ charge state. Differences in ATDs are calculated by subtracting the ATDs for metal coordinated octasaccharides from those of the metal free species. ATDs of calcium and potassium coordinated octasaccharides that exhibit two ion populations in the ion mobility spectrum are shown with the most abundant ion population represented. Figure taken from Seo *et al.*^[81]

Along with monitoring the conformational dynamics of GAGs, IMS has been used to separate isomeric species. The Leary group have reported applications of TWIMS-MS² for the separation and the sequencing of two heparan sulfate (HS) isomers, facilitated by ¹H NMR.^[83] Two isomeric HS hexasaccharides, purified by size exclusion and strong anion-exchanged chromatography, both produced highly similar mass spectra and enzymatic digestion profiles. TWIMS separation and subsequent CID of mobility separated ions was able to distinguish these epimeric hexasaccharides, which differed only by the stereochemistry at a single position (GlcA *vs* IdoA). Miller *et al.* also demonstrated TWIMS separation and identification of six tagged DP 8 (degree of polymerization) heparan sulfate isomers differing only by their presence and absence of IdoA and GlcA at the hexuronic positions.^[84] Fully IdoA DP8 structures displayed a more extended and flexible conformational distribution compared to the fully GlcA octasaccharide. The authors suggest this is because GlcA is reported to preferentially adopt the rigid ⁴C₁ chair conformation, whilst IdoA adopts a dynamic mixture of ¹C₄, ⁴C₁ and ²S₀ conformations.^[85] Interestingly, changes of IdoA to GlcA and *vice versa* within specific locations along the DP8 chain caused conformational distortions, which could be separated by TWIMS (Figure 17).

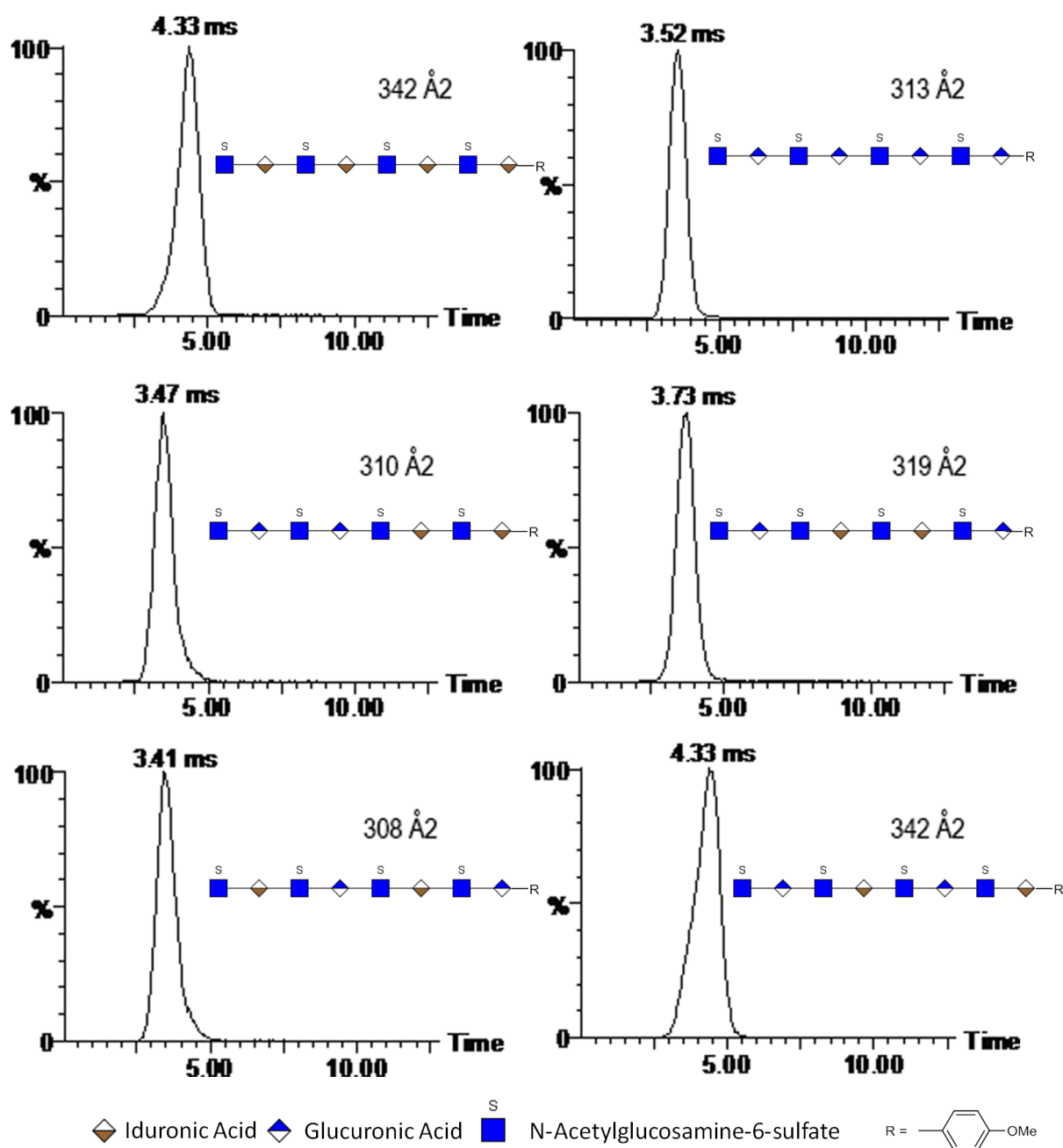


Figure 17 – Arrival time distributions and $^{TW}CCSN_2$ associated with six DP8 hexuronic acid positional isomers. Amended from Miller *et al.*^[84]

The intensity distribution of specific fragments differed when GlcA or IdoA was adjacent to the glycosidic bond cleavage site (more facile for GlcA) allowing unambiguous elucidation of the hexuronic acid sequence. Another report described the ability of IM-MS² to separate three synthetic heparin octasaccharides differing in sulfation or acetylation patterns employing a similar strategy.^[86] Two arrival time populations were observed for certain oligosaccharides, suggesting the existence of two distinct conformations bearing the same sulfation pattern. Indeed, the presence of sulfation at the 2-OH position of iduronic acid residues has been reported to lead to a mixture of conformers, corresponding to either a chair 1C_4 or/and a skew 2S_0 conformation.^[87] Recently, Lemr and co-workers

reported the use of TWIMS combined with MS² in order to separate hyaluronic acid isomers as well as to determine both their structural composition and their isomeric ratio.^[88] While IMS allowed separation of tyramine positional isomers, subsequent CID fragmentation of separated precursor ions led to similar mass fragments often with only differing intensities, making structural characterization challenging. To overcome this limitation, the reducing end of the GAG isomers were reductively labelled by NaBH₄ or stable isotope NABD₄ (both performed in H₂O), which increased the mass by 2 or 3 *m/z* respectively. This chemical modification resulted in the production of characteristic MS/MS fragments for each positional isomer and thus allowed their unambiguous identification and quantification even from mixtures.

Recently, Kailemia *et al.* have reported the first application of FAIMS, combined with Fourier transform ion cyclotron resonance mass spectrometry (FTICR-MS) and electron detachment dissociation (EDD) tandem MS, to mobility separate, select and characterize both anomeric and isobaric GAGs.^[89] A chondroitin sulfate A (CSA) mixture of oligomers at various degrees of polymerization (DP4 to DP10), which was prepared by partial enzymatic digestion, produced similar MS spectra. FAIMS was able to separate the mixture of oligomers at different degrees of polymerization, although each peak was not necessarily baseline resolved. Furthermore, some oligomers were observed in more than one peak, which was attributed to the presence of a mixture of reducing anomers. Synthetic HS tetramers, whose reducing anomeric alcohol had been methylated, locking the anomer as either α - or β -, both produced a single distinguishable ATD under identical FAIMS conditions suggesting this hypothesis may be true. Alternatively, chemical reduction of the free reducing ends using NaBH₄ to a single alditol species produced similar results. Mobility selected ions were fragmented by EDD, which is reported to preserve labile sulfate moieties,^[90] and *m/z* of the resultant product ions were measured on an FTICR-MS for high mass resolving power. While the fragment ions obtained in every sample were similar, different intensities were observed. PCA allowed differentiation of each of the stereoisomers.^[89]

Biologically relevant polysaccharides other than GAGs have also been studied by IMS to help separate isomers or monitor conformational variability. Rashid *et al.* have reported the discrimination of isobaric linear α -glucans with a DP of up to 35 using TWIMS in negative ion mode.^[33d] Several singly charged α 1-4-linked maltooligosaccharides and α 1-6-linked dextrans, with DP comprised between 2 and 10, were distinguishable by their ^{TW}CCS_{N₂}. Similarly, doubly charged and triply charged equivalents of the species between DP 3-24 and 14-35 respectively, were also distinguishable (Figure 18).

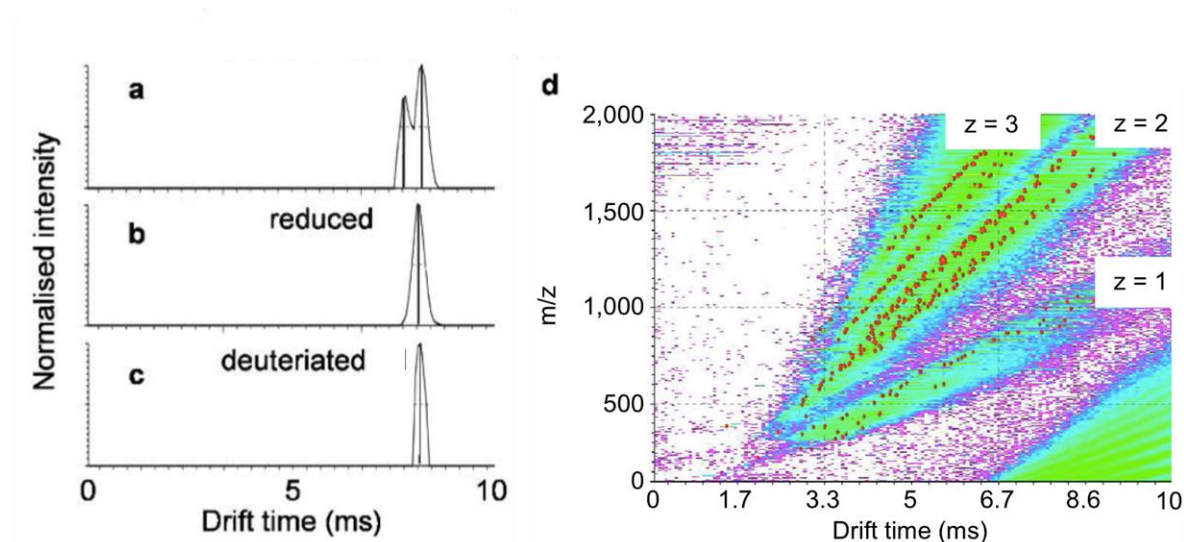


Figure 18 – Arrival time distribution associated with maltohexaose (a). The two species are thought to arise from a linear conformer and a V6 helical structure. After reduction (b) or deuteration (c), the conformational family reduces to a single, presumably disordered, species. Plot of drift time against m/z of a mixture of maltooligosaccharides dextran 5000 and amylose 2800 (maltooligosaccharides) showing singly, doubly and triply charged ion clouds (d). Red dots highlight significant ions. The trend line with a shorter drift time corresponds to amylose whereas the trend line with a longer drift time corresponds to dextran. Figure amended from Rashid *et al.*^[33d]

As might be expected, the mobility decreases with increasing DP or increasing oligomerization. Dextran, for the most part, exists as more extended conformers compared to the other equivalent DP polysaccharides, which is postulated to be as a result of the increased flexibility associated with the α 1-6 bond. The $^{TW}CCS_{N_2}$ did not linearly increase with increasing carbohydrate chain length, suggesting that the global conformation alters upon the successive addition of monomers. Nearly all the studied oligosaccharide ATDs consist of a single species, suggesting either mutarotation at the reducing terminus is minimal or anomers are inseparable under the conditions employed in this work. The only exception was the linear DP6 maltohexaose, which existed as two structures. However, only a single structure was observed after reduction and remarkably also after exchange of 18 of the 20 exchangeable hydrogen atoms (within the hydroxyl groups) with deuterium (Figure 18). This result was attributed to perturbation of the hydrogen bonding network within the maltohexaose. Maltooligosaccharides have been reported to adopt rigid left-handed V6 helices with six sugar units per turn in solution. Formation of the necessary intra-helical hydrogen-bonding interactions between the reducing and non-reducing termini of this V6 helix is probably dependent on the reducing residue mutarotation isoform.^[91] The structure of the DP6 sugar is therefore likely to be at the interface between existing as a linear-like chain and the V6 helical structure. Pullulans (α 1-4-linked maltotriose connected by α 1-6-linkages) were poorly separated from dextrans by IMS alone in agreement with solution-phase measurements where they are known to both adopt a similar random coil structure.^[92] However, these structures are readily distinguished by their CID tandem mass spectra. These studies enabled the characterization of the enzymatic product resulting from the action of *Mycobacterium tuberculosis* GlgB, a potential anti-tuberculosis drug target, on maltooligosaccharides.^[33d] This

enzyme is known to transfer a portion of linear maltooligosaccharide onto the 6-OH position of the maltooligosaccharide backbone forming conformationally extended, branched structures. The same year, Huang *et al.* have reported the IMS separation and MS² characterization of 3-aminoquinoline labelled isomeric oligogalacturonic acids (derived from pectin cell wall) in a mixture of oligosaccharides bearing various degrees of polymerization and levels of methyl esterification including certain methyl ester regioisomers.^[93] This suggests specific sites of methyl esterification affect the conformation more than others. However, there were instances where certain oligogalacturonic species were inseparable. In order to improve the separation of the isomeric galacturonic acid residues, the same group reported on-line hydrophilic interaction chromatography (HILIC) prior to TWIMS.^[94] HILIC-TWIMS-MS² allowed improved separation and discrimination of isomers. However, even using both HILIC and TWIMS separation, some isomers still remained extremely challenging to resolve. Estimations of the theoretical CCS_{N2} values highlighted that the current resolving power of TWIMS is insufficient to separate all isomeric methyl esterified oligogalacturonic acids completely. Other complex mixtures of oligosaccharides from a digest of sugar beet pectin have been subsequently studied, since they contain a large variety of both unknown structures and potential isomers. The combination of HILIC with TWIMS enabled simultaneous separation and characterization of isomeric beet rhamnogalacturonan I derived oligosaccharides based on the position of their glucuronic acid substituents. However, again, all isomers were not resolved using IMS alone due to the small differences in their ^{TW}CCS_{N2} values. Nevertheless, some novel structural isomers were identified for the first time, which demonstrated that *i*) oligosaccharide structures from beet pectin digest were more complex than previously reported and *ii*) IMS is suitable for the discovery of new structures.

Finally, in the recent study of Plancot *et al.*, IM-MS approaches were able to characterize arabinoxylan fragments obtained after *endo*-xylanase digestion of leaf extracts from two African plant species *Eragrostis nindensis* and *Eragrostis tef*.^[95] Multiple species were observed within the ATD suggesting the existence of structural isomers, confirmed by post-mobility tandem MS. Based on IM-MS/MS experiments for a DP4 oligosaccharide, the isomer at the shorter drift times could be assigned to a branched arabinoxylan structure and the one at the longer drift times to the linear one (Figure 19). However, although the presence of isomers was evidenced by IMS, fragments having higher DP were unresolvable.

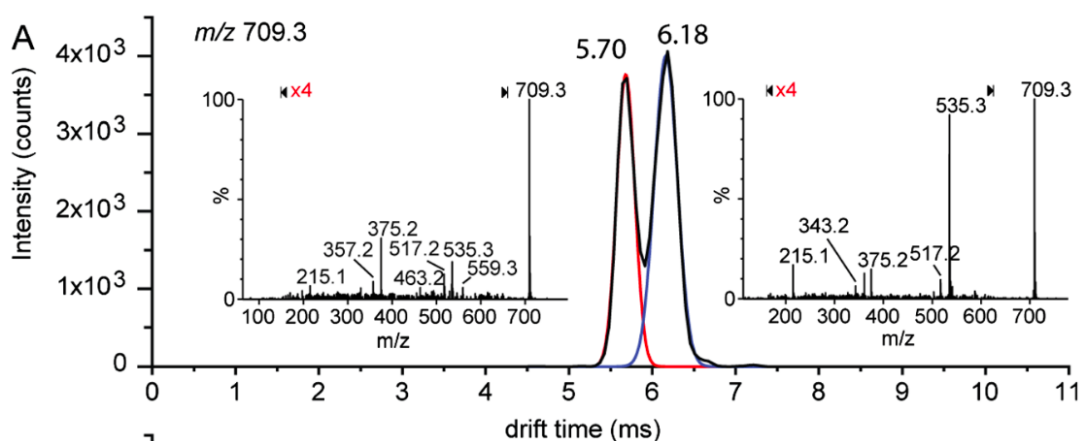


Figure 19 - Extracted ion mobility spectra of DP4 isomers (m/z 709). The insets show the tandem mass spectra associated with the drift times 5.70 and 6.18 ms. Figure taken from Plancot *et al.*^[95]

4.2 Glycopeptides/Glycoproteins

The complexity of analysing glycoproteins and glycopeptides as a single, intact entity is increased due the isomeric nature of many of the carbohydrate moieties. These factors have meant that MS techniques alone have often struggled to separate and therefore elucidate the different carbohydrate structures present and also define sites of glycosylation. With incorporation of IMS, analysis at the intact level has begun to emerge. The additional analytical dimension removes the requirement for lengthy deglycosylation and clean-up steps and in many cases has enabled simultaneous separation and identification.

Use of collisional activation or collision induced unfolding (CIU) has been reported during IMS experiments due to its ability to induce a change in conformation, identifiable by IMS. Leary studied the effects of increasing the trap collision energy of a TWIMS instrument on the ATDs and mobility profiles of the small glycoprotein, MCP-1.^[96] Monocyte chemotactic protein 1 (MCP-1) is a chemo-attractant cytokine that recruits monocytes, dendritic cells, and memory T cells to the sites of inflammation. MCP-1 exists in equilibrium between monomer, dimer and higher oligomeric forms; MCP-1 monomer activates its cognate receptors, while oligomerization and glycosaminoglycan (GAG) binding has been proven essential for *in vivo* activity. IMS was used to structurally probe the MCP-1 monomer, the non-covalent MCP-1 dimer and a dimer analogue non-covalently bound to a semi-synthetic GAG (Figure 20). With increasing collision energy, longer arrival times were observed for both dimers, indicative of a more extended conformation. However, the monomer species remained compact at equivalent activation energies. Comparison of the bound and unbound dimers revealed that GAG binding stabilized the dimer complex towards dissociation, indicating that the interaction was across the two MCP-1 monomers. In a second paper by the same group, IMS was used to show different structural specificity for GAG binding to two different chemokine variations. The interactions of chemokines with GAG moieties are essential for regulating chemokine

oligomerization, cell migration and cellular recruitment and so an understanding of how the overall conformation is affected upon binding is sought after.^[97] CIU has also been explored recently by the Ruotolo group for the larger, more complex system of a monoclonal antibody (mAb).^[98] Through use of collision energy ramps, mobility acquisition and heat map plots, the authors were able to demonstrate how the levels of glycosylation affect the unfolding of the mAb.

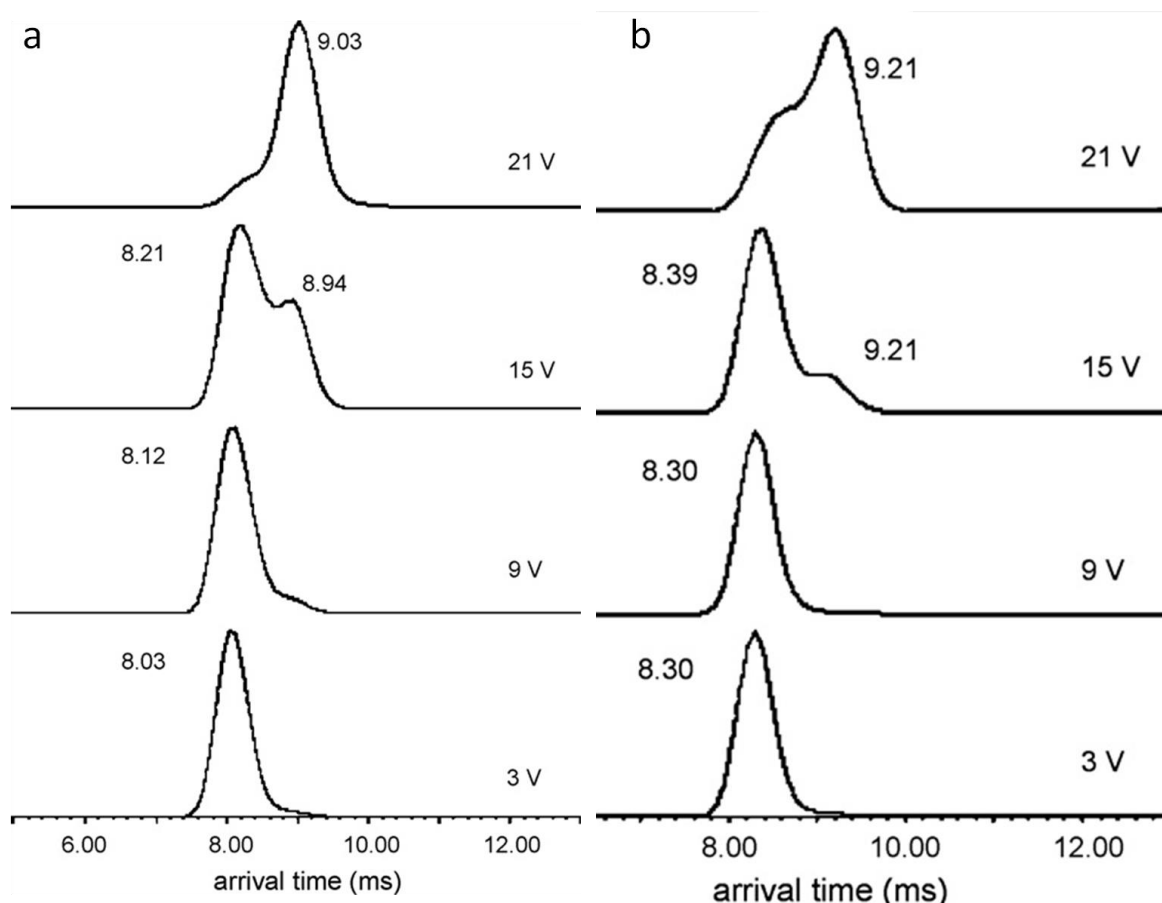


Figure 20 - The arrival time distributions (ATD) associated with the 9+ MCP-1 dimer, isolated in the quadrupole region, at the labelled collision energies 3, 9, 15, 21 and 27 V (a). Binding of the semi-synthetic GAG analogue Arixta is observed to stabilize the dimer at higher collision energies, although results in a less compact conformation (b). Figure amended from Schenauer *et al.*^[96]

DTIMS has also been applied by the Barran group to calculate $^{DT}CCS_{He}$ values for beta-defensins as free peptides and also for the glycoprotein equivalents; beta-defensins bound to heparin-derived disaccharides (HDD).^[99] The authors observed a 5% increase in $^{DT}CCS_{He}$ for the bound form signifying a strong interaction with a defined structure, further evidenced by the narrower ATD compared with the unbound form. Molecular modelling provided CCS_{He} values for comparison, although in all cases the $^{DT}CCS_{He}$ values for bound HDD were less than expected. The conformations observed indicate that linear defensins may still access a functional fold. Similar techniques were applied to examine the interaction of metamorphic protein lymphotactin with fondaparinux. Upon titration of the carbohydrate with the protein, rapid aggregation was observed, which resulted in μm

assemblies.^[100] Mutated and truncated forms of the protein formed complexes of different stoichiometries with distinctive $^{DT}CCS_{He}$. These studies highlight how combining predictive methodologies such as molecular modelling with structural techniques such as IM can identify functional forms of proteins and interactions that can help to discern mode of action as well the formation of macromolecular structures.

In addition to structural information, IMS enables separation of isomeric structures with the same m/z . Li *et al.* used a trend line separation method coupled with IMS to dramatically increase the signal-to-noise ratio of glycopeptides derived from a glycoprotein using trypsin digestion.^[101] One of the glycoproteins analysed has a significant carbohydrate content of approximately 45% across 5 N-glycosylation sites. With such a high carbohydrate content, the likelihood of isomeric carbohydrates and hence isomeric or stereochemical glycopeptides is high. For some glycopeptides, multiple features in the ATD were observed for a single m/z emphasizing the ability of IMS to distinguish between isomeric glycopeptides.^[101] Separation of isomeric glycopeptides, differing only by the site of glycosylation, has also been reported by Creese and Cooper using FAIMS (Figure 21).^[102] Combining FAIMS with routine bottom-up proteomic strategies has been shown to facilitate comprehensive mapping of the O-glycosylation profile of a glycoprotein, including identification of novel glycans and glycosylation sites.^[103]

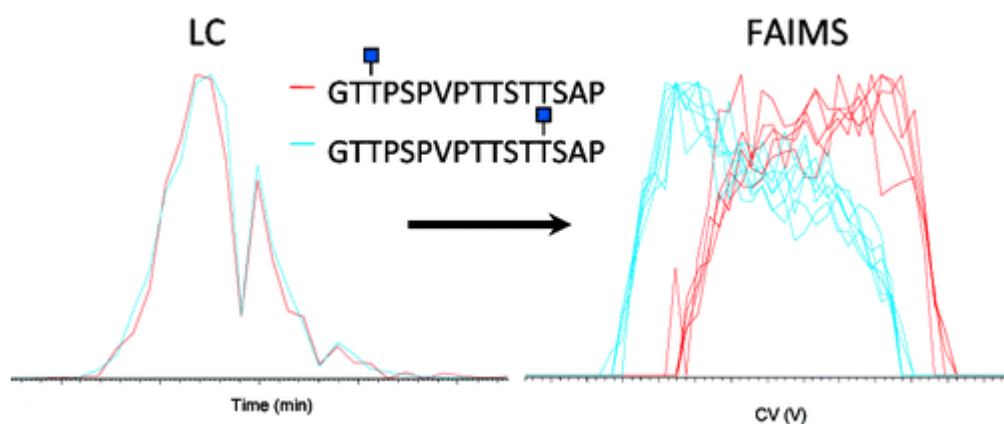


Figure 21 – LC chromatogram highlighting lack of separation of two glycopeptides differing only by their site of glycosylation and the equivalent FAIMS trace where they are distinguishable. Figure taken from Creese.^[102]

Analysis of bound glycoproteins and glycopeptides without substantial deglycosylation and/or digestion and/or purification is still relatively unexplored due to the challenges faced by MS techniques being unable to distinguish between the isomeric forms of the carbohydrates and their linked locations. However, incorporation of IMS has offered an additional dimension, allowing for the separation and identification of glycans without the need for removal from the peptide backbone. An advantageous side effect of these analyses is that structural characteristics at the intact level are observable and can offer hypotheses for functionality requirements.

4.3 Glycolipids

Glycolipids are amphiphilic constituents of cell membranes containing a hydrophilic carbohydrate, varying from simple monosaccharides to complex polysaccharides, covalently bound to a hydrophobic lipid moiety. The growing interest in profiling cellular glycolipids has been motivated by the importance of the carbohydrate head groups in many biological processes including recognition events, adhesion and signalling pathways as well as their architectural role in cell membranes.^[104] Among the glycolipid family, lipopolysaccharides (LPS) from bacterial origin may modulate the immune response and have been extensively used as components of antibacterial vaccines against a large variety of bacteria such as *Haemophilus influenzae* type b,^[105] *Neisseria meningitidis* type C^[106] and *Streptococcus pneumoniae*.^[107] Animals also contain the more complex glycosphingolipids, especially within the central nervous system, where the glycan part is linked to a ceramide moiety (composed of a sphingosine and a fatty acid).^[108] They are also reported to be vital for brain development, neuritogenesis, synaptic transmission and aging^[109] and also for many diseases such as Alzheimer's, Huntington's, Parkinson's, Niemann-Pick type C, Gaucher's and Guillain-Barré syndrome.^[110] Moreover, aberrant expressions of certain glycosphingolipids have been linked to several types of cancer, and hence, they have been frequently used as diagnostic biomarkers for different stages of cancer.^[111] Because of both their biological relevant functions and promising medical application, it seems essential to well characterize this family of glycoconjugates. However, analysis of glycolipids is not a simple task due to the inherent heterogeneity and complexity of their structure including the nature, number and position of the different monosaccharide residues, as well as the length, saturation and configuration of the carbon chains of lipid moieties.

Li and co-workers were the first to report the combination of capillary electrophoresis (CE) and FAIMS-MS in order to analyse a mixture of O-deacylated LPS from *Haemophilus influenzae* strain 375.^[112] Although the potential of CE-FAIMS-MS to separate a complex mixture of LPS was demonstrated, FAIMS was solely used in this study to 'filter out' the chemical background, thus enhancing detection limits. In 2007, Woods *et al.* studied the spatial distribution of cerebrosides species in rat brain section using MALDI-MS coupled to IMS.^[113] A few years later, Shvartsburg *et al.* described the separation of a variety of lipids using FAIMS. They have shown that monogalactosyl diacylglycerols (MGDG) and digalactosyl diacylglycerols (DGDG) fall on the same trend line within a 2D plot of compensation voltage and m/z and thus their MS information could be readily isolated from a complex mixture of lipids. The same year, Woods and co-workers reported the use of MALDI-IM-TOF to separate a mixture of gangliosides, a well-studied class of glycosphingolipids, containing different proportions of negatively charged sialic acid (*N*-acetylneuraminic acid) as caesium adducts. Although two purified structural isomers GD1a and GD1b, which only differ by the position of the two sialic acids, generated identical MS spectra, GD1a had a greater mobility compared to GD1b. IMS was able to separate, based on their degree of sialylation (*i.e.* GM1, GD1 and GT1), several gangliosides extracted from a mouse cerebrum, although the structural isomers GD1a and GD1b were poorly resolved in most cases.^[114] In a recent study, the group of Woods also reported the analysis of

similar isomers using IM-MS in negative ion mode circumventing the need for doping the sample with caesium salts, which would otherwise suppress the intensity of the studied glycolipids.^[115] Signal is also suppressed by the presence of phosphatidylcholines and sphingomyelins, which both contain quaternary amines and so ionize very well in positive ion mode. However, even in negative ion mode the GD1 isomers were inseparable based upon their mobility.^[115] Notable trends were observed for the effect of lipid structure upon mobility for the negatively charged species akin to those mentioned for salt adducts.^[116] Reducing the head size, increasing the radyl chain length, increasing degree of saturation, and increasing the number of sialic acids all increase the mobility.^[115]

5. Concluding Remarks and Future Outlook

Over the past decade, the major advances in MS-based glycomic and glycoproteomic techniques, as well as chromatographic separation methods, have allowed identification of subtle structural changes associated with disease development and progression.^[117] However, so far, only a few reports have described the use of IM-MS to perform this task especially for glycomic applications, where additional separation technologies and structural information are urgently needed.^[10d, 10e, 62, 118] The resolving power of DTIMS,^[52a] TWIMS^[30] and FAIMS^[119] have improved significantly over a relatively short period of time, addressing initial issues of poor separation of certain isomeric carbohydrates. Also alternative forms of mobility separation are being developed, which show significant advances in resolving power. Kurulugama *et al.* developed a strategy termed overtone mobility spectrometry (OMS), which yields significantly improved resolving powers than the DTIMS from which it is derived. This instrument consists of a segmented drift tube, to which a uniform potential is applied across segment(s) at regular intervals forming a ‘sawtooth’ gradient along the mobility cell. These phases are alternatively switched on and off and ions are only transmitted when their mobilities are in resonance with the phase change frequency. An interesting result is that overtones are observed.^[50a, 120] Cyclotron geometry segmented drift cell have also been developed,^[121] where ion packets are propelled around the tube by changing the drift field at a frequency resonant with the ions drift time through each region of the cyclotron. Ions out of resonance are not stable; therefore the cyclotron acts to trap ions of a given mobility. Increasing the number of cycles improves the resolving power (>300 for more than 10 cycles and >1000 for 100 cycles).^[121b, 121c] The downside of this is increased ion loss as the number of cycles is increased and a reduction of duty cycle, both of which reduce the sensitivity. Either way, improvement and optimization of this approach or development of novel technologies, will see the resolving power significantly improved with sufficient sensitivity and speed for analysis of biological glycans. Improvements associated with the speed, structure generation and usability of computational simulations (*i.e. ab initio*, semi-empirical or molecular dynamics) to generate theoretical candidate structures and CCSs will also facilitate

validation and characterization of dynamic structural/conformational information measured by IM-MSⁿ.

Given that the structure, presence and proportion of specific glycan-biomarkers can change during the course of disease and these structures also vary from individual-to-individual, there will undoubtedly be benefits in developing glycan-based diagnostic tools for personalized medicine. As a result, rapid, sensitive and robust methods to characterize and identify biological glycans are required in order to carry out both diagnosis and prognosis, especially approaches that can detect diseases early. In fact, structural identification of aberrant glycosylation or specific structures associated with disease have already proved beneficial in the development of a large number of carbohydrate-based vaccines over the last 20 years such as anticancer,^[122] antibacterial^[123] or antiviral^[124] with many more still in development.^[125] There will also be the requirement to assess the quality of any isolated or synthesized carbohydrate-based medicines, further driving the requirement of robust analytical techniques. Initial applications of IM-MS to facilitate separation and identification of glycans,^[33c, 48a, 50b, 56, 74, 126] glycosides,^[19b, 62, 114-115] polysaccharides,^[33f, 84, 86] glyco(peptides/proteins)^[102] and glycan-based biomarkers^[10c-e, 62] are extremely promising, and further development and the recent commercialization of IMS technologies will inevitably increase its use within research and healthcare. The development of universal sequencing tools for carbohydrates with comparable speed and throughput to those used for nucleic acids and proteins remains a challenge, and IM-MS techniques are promising to be useful tools in this context.

Acknowledgements

We are grateful to the BBSRC (studentship to CJG, RU, LM and funding for the IBCarb network), Covance Laboratories (icase to RU) the FP7 programme (GlycoBioM, Immunoshape, GLYCOVAX) and the Royal Society (Wolfson Merit award to SLF) for supporting our work.

References

- [1] K. Ohtsubo, J. D. Marth, *Cell* **2006**, 126, 855.
- [2] a)T. Yoshida-Moriguchi, L. P. Yu, S. H. Stalnakker, S. Davis, S. Kunz, M. Madson, M. B. A. Oldstone, H. Schachter, L. Wells, K. P. Campbell, *Science* **2010**, 327, 88; b)M. Phillips, E. Nudelman, F. Gaeta, M. Perez, A. Singhal, S. Hakomori, J. Paulson, *Science* **1990**, 250, 1130.
- [3] a)L. Bode, *Glycobiology* **2012**; b)M. B. Poulin, Y. Shi, C. Protsko, S. A. Dalrymple, D. A. R. Sanders, B. M. Pinto, T. L. Lowary, *Chembiochem* **2014**, 15, 47; c)T. Boren, P. Falk, K. A. Roth, G. Larson, S. Normark, *Science* **1993**, 262, 1892.
- [4] M. Sperandio, C. A. Gleissner, K. Ley, *Immunological Reviews* **2009**, 230, 97.
- [5] a)H. R. Morris, A. Dell, R. L. Easton, M. Panico, H. Koistinen, R. Koistinen, S. Oehninger, M. S. Patankar, M. Seppala, G. F. Clark, *Journal of Biological Chemistry* **1996**, 271, 32159; b)S. Benoff, *Molecular Human Reproduction* **1997**, 3, 599; c)S. Schröter, C. Osterhoff, W. McArdle, R. Ivell, *Human Reproduction Update* **1999**, 5, 302.

- [6] a)J. Bones, S. Mittermayr, N. O'Donoghue, A. Guttman, P. M. Rudd, *Analytical Chemistry* **2010**, 82, 10208; b)G. Lauc, J. E. Huffman, M. Pucic, L. Zgaga, B. Adamczyk, A. Muzinic, M. Novokmet, O. Polasek, O. Gornik, J. Kristic, T. Keser, V. Vitart, B. Scheijen, H. W. Uh, M. Molokhia, A. L. Patrick, P. McKeigue, I. Kolcic, I. K. Lukic, O. Swann, F. N. van Leeuwen, L. R. Ruhaak, J. J. Houwing-Duistermaat, P. E. Slagboom, M. Beekman, A. J. M. de Craen, A. M. Deelder, Q. Zeng, W. Wang, N. D. Hastie, U. Gyllensten, J. F. Wilson, M. Wuhrer, A. F. Wright, P. M. Rudd, C. Hayward, Y. Aulchenko, H. Campbell, I. Rudan, *Plos Genetics* **2013**, 9; c)T. Iskratsch, A. Braun, K. Paschinger, I. B. H. Wilson, *Analytical Biochemistry* **2009**, 386, 133; d)S. R. Kumar, E. R. Sauter, T. P. Quinn, S. L. Deutscher, *Clinical Cancer Research* **2005**, 11, 6868; e)R. M. Anthony, F. Nimmerjahn, D. J. Ashline, V. N. Reinhold, J. C. Paulson, J. V. Ravetch, *Science* **2008**, 320, 373; f)Y. Kanda, T. Yamada, K. Mori, A. Okazaki, M. Inoue, K. Kitajima-Miyama, R. Kuni-Kamochi, R. Nakano, K. Yano, S. Kakita, K. Shitara, M. Satoh, *Glycobiology* **2007**, 17, 104; g)G. Thanabalasingham, J. E. Huffman, J. J. Kattla, M. Novokmet, I. Rudan, A. L. Gloyn, C. Hayward, B. Adamczyk, R. M. Reynolds, A. Muzinic, N. Hassanali, M. Pucic, A. J. Bennett, A. Essafi, O. Polasek, S. A. Mughal, I. Redzic, D. Primorac, L. Zgaga, I. Kokic, T. Hansen, D. Gasperikova, E. Tjora, M. W. J. Strachan, T. Nielsen, J. Stanik, I. Klimes, O. B. Pedersen, P. R. Njolstad, S. H. Wild, U. Gyllensten, O. Gornik, J. F. Wilson, N. D. Hastie, H. Campbell, M. I. McCarthy, P. M. Rudd, K. R. Owen, G. Lauc, A. F. Wright, *Diabetes* **2013**, 62, 1329; h)H. F. Lodish, A. Berk, S. Zipursky, P. Matsudaira, D. Baltimore, J. Darnell, *Molecular Cell Biology*, 4th edition ed., New York, **2000**.
- [7] M. Wuhrer, C. A. M. Koeleman, J. M. Fitzpatrick, K. F. Hoffmann, A. M. Deelder, C. H. Hokke, *Glycobiology* **2006**, 16, 991.
- [8] F. Yamamoto, *Immunohematology / American Red Cross* **2004**, 20, 3.
- [9] a)J. Krištić, F. Vučković, C. Menni, L. Klarić, T. Keser, I. Beceheli, M. Pučić-Baković, M. Novokmet, M. Mangino, K. Thaqi, P. Rudan, N. Novokmet, J. Šarac, S. Missoni, I. Kolčić, O. Polašek, I. Rudan, H. Campbell, C. Hayward, Y. Aulchenko, A. Valdes, J. F. Wilson, O. Gornik, D. Primorac, V. Zoldoš, T. Spector, G. Lauc, *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences* **2014**, 69, 779; b)I. Gudelj, T. Keser, F. Vuckovic, V. Skaro, S. S. Goreta, T. Pavic, J. Dumic, D. Primorac, G. Lauc, O. Gornik, *International Journal of Legal Medicine* **2015**, 129, 955; c)L. Krishnamoorthy, L. K. Mahal, *Acs Chemical Biology* **2009**, 4, 715.
- [10] a)O. Gornik, I. Gornik, V. Gasparovic, G. Lauc, *Clinical Biochemistry* **2008**, 41, 504; b)I. T. Akmacic, N. Ventham, E. Theodoratou, F. Vuckovic, N. Kennedy, J. Kristic, E. R. Nimmo, R. Kalla, H. Drummond, J. Stambuk, M. G. Dunlop, M. Novokmet, Y. Aulchenko, O. Gornik, H. Campbell, M. Pucic-Bakovic, J. Satsangi, G. Lauc, I. B. Consortium, *Journal of Crohns & Colitis* **2015**, 9, S107; c)D. Isailovic, R. T. Kurulugama, M. D. Plasencia, S. T. Stokes, Z. Kyselova, R. Goldman, Y. Mechref, M. V. Novotny, D. E. Clemmer, *Journal of Proteome Research* **2008**, 7, 1109; d)D. Isailovic, M. D. Plasencia, M. M. Gaye, S. T. Stokes, R. T. Kurulugama, V. Pungpapong, M. Zhang, Z. Kyselova, R. Goldman, Y. Mechref, M. V. Novotny, D. E. Clemmer, *Journal of Proteome Research* **2012**, 11, 576; e)M. M. Gaye, S. J. Valentine, Y. Hu, N. Mirjankar, Z. T. Hammoud, Y. Mechref, B. K. Lavine, D. E. Clemmer, *Journal of Proteome Research* **2012**, 11, 6102.
- [11] a)K. Ohtsubo, S. Takamatsu, M. T. Minowa, A. Yoshida, M. Takeuchi, J. D. Marth, *Cell* **2005**, 123, 1307; b)C. R. Varki A, Esko JD, et al., *Essentials of Glycobiology*, 2 ed., Cold Spring Harbor, New York, **2009**.
- [12] L. Royle, A. Roos, D. J. Harvey, M. R. Wormald, D. Van Gijlswijk-Janssen, E.-R. M. Redwan, I. A. Wilson, M. R. Daha, R. A. Dwek, P. M. Rudd, *Journal of Biological Chemistry* **2003**, 278, 20140.
- [13] a)E. Gorelik, U. Galili, A. Raz, *Cancer metastasis reviews* **2001**, 20, 245; b)M. J. Paszek, C. C. DuFort, O. Rossier, R. Bainer, J. K. Mouw, K. Godula, J. E. Hudak, J. N. Lakins, A. C. Wijekoon, L. Cassereau, M. G. Rubashkin, M. J. Magbanua, K. S. Thorn, M. W. Davidson, H. S. Rugo, J. W. Park, D. A. Hammer, G. Giannone, C. R. Bertozzi, V. M. Weaver, *Nature* **2014**, 511, 319; c)F. Jacob, D. R. Goldstein, N. V. Bovin, T. Pochechueva, M. Spengler, R. Caduff, D. Fink, M. I. Vuskovic, M. E. Huflejt, V. Heinzelmann-Schwarz, *International*

- Journal of Cancer* **2012**, 130, 138; d)G. F. Springer, *Science* **1984**, 224, 1198; e)Y. T. Chen, Y. M. Chong, C. W. Cheng, C. L. Ho, H. W. Tsai, F. H. Kasten, Y. L. Chen, C. F. Chang, *Clinica Chimica Acta* **2013**, 420, 45.
- [14] D. M. Krasnewich, G. D. Holt, M. Brantly, F. Skovby, J. Redwine, W. A. Gahl, *Glycobiology* **1995**, 5, 503.
- [15] F. Vuckovic, J. Kristic, I. Gudelj, M. Teruel, T. Keser, M. Pezer, M. Pucic-Bakovic, J. Stambuk, I. Trbojevic-Akmacic, C. Barrios, T. Pavic, C. Menni, Y. X. Wang, Y. Zhou, L. F. Cui, H. C. Song, Q. Zeng, X. H. Guo, B. A. Pons-Estel, P. McKeigue, A. L. Patrick, O. Gornik, T. D. Spector, M. Harjacek, M. Alarcon-Riquelme, M. Molokhia, W. Wang, G. Lauc, *Arthritis & Rheumatology* **2015**, 67, 2978.
- [16] a)A. Weidemann, G. König, D. Bunke, P. Fischer, J. M. Salbaum, C. L. Masters, K. Beyreuther, *Cell* **1989**, 57, 115; b)P. Pahlsson, S. L. Spitalnik, *Archives of Biochemistry and Biophysics* **1996**, 331, 177.
- [17] a)J. Clardy, C. Walsh, *Nature* **2004**, 432, 829; b)M. Dalziel, M. Crispin, C. N. Scanlan, N. Zitzmann, R. A. Dwek, *Science* **2014**, 343; c)M. N. Thaker, G. D. Wright, *Acs Synthetic Biology* **2015**, 4, 195.
- [18] a)L. Royle, M. P. Campbell, C. M. Radcliffe, D. M. White, D. J. Harvey, J. L. Abrahams, Y. G. Kim, G. W. Henry, N. A. Shadick, M. E. Weinblatt, D. M. Lee, P. M. Rudd, R. A. Dwek, *Analytical Biochemistry* **2008**, 376, 1; b)M. P. Campbell, L. Royle, C. M. Radcliffe, R. A. Dwek, P. M. Rudd, *Bioinformatics* **2008**, 24, 1214.
- [19] a)S. Itoh, N. Kawasaki, M. Ohta, M. Hyuga, S. Hyuga, T. Hayakawa, *Journal of Chromatography A* **2002**, 968, 89; b)M. Pičmanová, E. H. Neilson, M. S. Motawia, C. E. Olsen, N. Agerbirk, C. J. Gray, S. Flitsch, S. Meier, D. Silvestro, K. Jørgensen, R. Sánchez Pérez, B. L. Møller, N. Bjarnholt, *Biochemical Journal* **2015**, 469, 375; c)L. R. Ruhaak, A. M. Deelder, M. Wührer, *Analytical and Bioanalytical Chemistry* **2009**, 394, 163.
- [20] a)C. A. Cooper, E. Gasteiger, N. H. Packer, *Proteomics* **2001**, 1, 340; b)H. L. Zhang, S. Singh, V. N. Reinhold, *Analytical Chemistry* **2005**, 77, 6263; c)C. A. Cooper, M. J. Harrison, M. R. Wilkins, N. H. Packer, *Nucleic Acids Research* **2001**, 29, 332; d)O. M. Saad, J. A. Leary, *Analytical Chemistry* **2005**, 77, 5902; e)E. P. Go, K. R. Rebecchi, D. S. Dalpathado, M. L. Bandu, Y. Zhang, H. Desaire, *Analytical Chemistry* **2007**, 79, 1708; f)C. A. Hayes, N. G. Karlsson, W. B. Struwe, F. Lisacek, P. M. Rudd, N. H. Packer, M. P. Campbell, *Bioinformatics* **2011**, 27, 1343; g)M. Frank, T. Lutteke, C. W. von der Lieth, *Nucleic Acids Research* **2007**, 35, D287; h)M. P. Campbell, T. Nguyen-Khuong, C. A. Hayes, S. A. Flowers, K. Alagesan, D. Kolarich, N. H. Packer, N. G. Karlsson, *Biochimica Et Biophysica Acta-Proteins and Proteomics* **2014**, 1844, 108; i)C. Konda, F. A. Londry, B. Bendiak, Y. Xia, *Journal of the American Society for Mass Spectrometry* **2014**, 25, 1441.
- [21] B. Domon, C. E. Costello, *Glycoconjugate Journal* **1988**, 5, 397.
- [22] a)C. S. Creaser, J. R. Griffiths, C. J. Bramwell, S. Noreen, C. A. Hill, C. L. P. Thomas, *Analyst* **2004**, 129, 984; b)F. Lanucara, S. W. Holman, C. J. Gray, C. E. Eyers, *Nat Chem* **2014**, 6, 281.
- [23] a)C. Uetrecht, R. J. Rose, E. van Duijn, K. Lorenzen, A. J. R. Heck, *Chemical Society Reviews* **2010**, 39, 1633; b)A. B. Kanu, P. Dwivedi, M. Tam, L. Matz, H. H. Hill, *Journal of Mass Spectrometry* **2008**, 43, 1.
- [24] a)T. Wytenbach, N. A. Pierson, D. E. Clemmer, M. T. Bowers, in *Annual Review of Physical Chemistry, Vol 65, Vol. 65* (Eds.: M. A. Johnson, T. J. Martinez), **2014**, pp. 175; b)V. D'Atri, M. Porrini, F. Rosu, V. Gabelica, *Journal of Mass Spectrometry* **2015**, 50, 711.
- [25] a)E. A. Mason, H. W. Schamp, *Annals of Physics* **1958**, 4, 233; b)R. T. Kurulugama, E. Darland, F. Kuhlmann, G. Stafford, J. Fjeldsted, *Analyst* **2015**, 140, 6834.
- [26] a)H. W. Ellis, R. Y. Pai, E. W. McDaniel, E. A. Mason, L. A. Viehland, *Atomic Data and Nuclear Data Tables* **1976**, 17, 177; b)H. W. Ellis, E. W. McDaniel, D. L. Albritton, L. A. Viehland, S. L. Lin, E. A. Mason, *Atomic Data and Nuclear Data Tables* **1978**, 22, 179.
- [27] a)H. W. Ellis, M. G. Thackston, E. W. McDaniel, E. A. Mason, *Atomic Data and Nuclear Data Tables* **1984**, 31, 113; b)L. A. Viehland, E. A. Mason, *Atomic Data and Nuclear Data Tables* **1995**, 60, 37.

- [28] J. C. May, C. R. Goodwin, N. M. Lareau, K. L. Leaptrot, C. B. Morris, R. T. Kurulugama, A. Mordehai, C. Klein, W. Barry, E. Darland, G. Overney, K. Imatani, G. C. Stafford, J. C. Fjeldsted, J. A. McLean, *Analytical Chemistry* **2014**, 86, 2107.
- [29] K. Giles, S. D. Pringle, K. R. Worthington, D. Little, J. L. Wildgoose, R. H. Bateman, *Rapid Communications in Mass Spectrometry* **2004**, 18, 2401.
- [30] K. Giles, J. P. Williams, I. Campuzano, *Rapid Communications in Mass Spectrometry* **2011**, 25, 1559.
- [31] a)A. S. Gelb, R. E. Jarratt, Y. T. Huang, E. D. Dodds, *Analytical Chemistry* **2014**, 86, 11396; b)J. Hofmann, W. B. Struwe, C. A. Scarff, J. H. Scrivens, D. J. Harvey, K. Pagel, *Analytical Chemistry* **2014**, 86, 10789.
- [32] C. Bleiholder, N. R. Johnson, S. Contreras, T. Wyttenbach, M. T. Bowers, *Analytical Chemistry* **2015**, 87, 7196.
- [33] a)M. F. Jarrold, *Physical Chemistry Chemical Physics* **2007**, 9, 1659; b)E. W. Robinson, E. R. Williams, *Journal of the American Society for Mass Spectrometry* **2005**, 16, 1427; c)J. P. Williams, M. Grabenauer, R. J. Holland, C. J. Carpenter, M. R. Wormald, K. Giles, D. J. Harvey, R. H. Bateman, J. H. Scrivens, M. T. Bowers, *International Journal of Mass Spectrometry* **2010**, 298, 119; d)A. M. Rashid, G. Saalbach, S. Bornemann, *Rapid Communications in Mass Spectrometry* **2014**, 28, 191; e)L. Jin, P. E. Barran, J. A. Deakin, M. Lyon, D. Uhrin, *Physical Chemistry Chemical Physics* **2005**, 7, 3464; f)Y. J. Seo, M. R. Schenauer, J. A. Leary, *International Journal of Mass Spectrometry* **2011**, 303, 191; g)B. T. Ruotolo, K. Giles, I. Campuzano, A. M. Sandercock, R. H. Bateman, C. V. Robinson, *Science* **2005**, 310, 1658; h)B. T. Ruotolo, J. L. P. Benesch, A. M. Sandercock, S. J. Hyung, C. V. Robinson, *Nature Protocols* **2008**, 3, 1139.
- [34] R. Cumeras, E. Figueras, C. E. Davis, J. I. Baumbach, I. Gracia, *Analyst* **2015**, 140, 1376.
- [35] R. W. Purves, R. Guevremont, *Analytical Chemistry* **1999**, 71, 2346.
- [36] A. A. Shvartsburg, T. A. Seim, W. F. Danielson, R. Norheim, R. J. Moore, G. A. Anderson, R. D. Smith, *Journal of the American Society for Mass Spectrometry* **2013**, 24, 109.
- [37] R. Guevremont, *Journal of Chromatography A* **2004**, 1058, 3.
- [38] A. A. Shvartsburg, Y. M. Ibrahim, R. D. Smith, *Journal of the American Society for Mass Spectrometry* **2014**, 25, 480.
- [39] a)F. Fernandez-Lima, D. Kaplan, J. Suetering, M. Park, *International Journal for Ion Mobility Spectrometry* **2011**, 14, 93; b)K. Michelmann, J. Silveira, M. Ridgeway, M. Park, *Journal of the American Society for Mass Spectrometry* **2015**, 26, 14.
- [40] J. A. Silveira, M. E. Ridgeway, M. A. Park, *Analytical Chemistry* **2014**, 86, 5624.
- [41] S. Koeniger, D. Clemmer, *Journal of the American Society for Mass Spectrometry* **2007**, 18, 322.
- [42] a)E. Mack, *Journal of the American Chemical Society* **1925**, 47, 2468; b)G. Vonhelden, M. T. Hsu, N. Gotts, M. T. Bowers, *Journal of Physical Chemistry* **1993**, 97, 8182.
- [43] M. F. Mesleh, J. M. Hunter, A. A. Shvartsburg, G. C. Schatz, M. F. Jarrold, *Journal of Physical Chemistry* **1996**, 100, 16082.
- [44] a)C. Bleiholder, T. Wyttenbach, M. T. Bowers, *International Journal of Mass Spectrometry* **2011**, 308, 1; b)S. E. Anderson, C. Bleiholder, E. R. Brocker, P. J. Stang, M. T. Bowers, *International Journal of Mass Spectrometry* **2012**, 330, 78; c)C. Bleiholder, S. Contreras, M. T. Bowers, *International Journal of Mass Spectrometry* **2013**, 354, 275; d)C. Bleiholder, S. Contreras, T. D. Do, M. T. Bowers, *International Journal of Mass Spectrometry* **2013**, 345, 89; e)T. Wyttenbach, C. Bleiholder, S. E. Anderson, M. T. Bowers, *Molecular Physics* **2015**, 113, 2344.
- [45] a)A. A. Shvartsburg, M. F. Jarrold, *Chemical Physics Letters* **1996**, 261, 86; b)A. A. Shvartsburg, S. V. Mashkevich, E. S. Baker, R. D. Smith, *Journal of Physical Chemistry A* **2007**, 111, 2002; c)C. Larriba-Andaluz, C. J. Hogan, Jr., *Journal of Chemical Physics* **2014**, 141.
- [46] I. Campuzano, M. F. Bush, C. V. Robinson, C. Beaumont, K. Richardson, H. Kim, H. I. Kim, *Analytical Chemistry* **2012**, 84, 1026.
- [47] L. S. Fenn, J. A. McLean, *Chemical Communications* **2008**, 5505.

- [48] a)J. Hofmann, H. S. Hahm, P. H. Seeberger, K. Pagel, *Nature* **2015**, 526, 241; b)P. Dwivedi, B. Bendiak, B. Clowers, H. Hill, *Journal of the American Society for Mass Spectrometry* **2007**, 18, 1163; c)W. Gabryelski, K. L. Froese, *Journal of the American Society for Mass Spectrometry* **2003**, 14, 265; d)D. J. Harvey, C. A. Scarff, M. Crispin, C. N. Scanlan, C. Bonomelli, J. H. Scrivens, *Journal of the American Society for Mass Spectrometry* **2012**, 23, 1955.
- [49] a)P. M. Lalli, Y. E. Corilo, M. Fasciotti, M. F. Riccio, G. F. de Sa, R. J. Daroda, G. H. M. F. Souza, M. McCullagh, M. D. Bartberger, M. N. Eberlin, I. D. G. Campuzano, *Journal of Mass Spectrometry* **2013**, 48, 989; b)G. A. Bataglione, G. Souza, G. Heerdt, N. H. Morgon, J. D. L. Dutra, R. O. Freire, M. N. Eberlin, A. Tata, *Journal of Mass Spectrometry* **2015**, 50, 336.
- [50] a)R. T. Kurulugama, F. M. Nachtigall, S. Lee, S. J. Valentine, D. E. Clemmer, *Journal of the American Society for Mass Spectrometry* **2009**, 20, 729; b)P. Both, A. P. Green, C. J. Gray, R. Šardžik, J. Voglmeir, C. Fontana, M. Austeri, M. Rejzek, D. Richardson, R. A. Field, G. Widmalm, S. L. Flitsch, C. E. Eyers, *Nat Chem* **2014**, 6, 65.
- [51] M. Fasciotti, G. B. Sanvido, V. G. Santos, P. M. Lalli, M. McCullagh, G. F. de Sá, R. J. Daroda, M. G. Peter, M. N. Eberlin, *Journal of Mass Spectrometry* **2012**, 47, 1643.
- [52] a)E. J. Davis, K. F. Grows, W. F. Siems, H. H. Hill, *Analytical Chemistry* **2012**, 84, 4858; b)J. C. May, J. N. Dodds, R. T. Kurulugama, G. C. Stafford, J. C. Fjeldsted, J. A. McLean, *Analyst* **2015**, 140, 6824.
- [53] L. S. Fenn, J. A. McLean, *Physical Chemistry Chemical Physics* **2011**, 13, 2196.
- [54] S. Lee, T. Wyttenbach, M. T. Bowers, *International Journal of Mass Spectrometry and Ion Processes* **1997**, 167–168, 605.
- [55] a)M. D. Leavell, S. P. Gaucher, J. A. Leary, J. A. Taraszka, D. E. Clemmer, *Journal of the American Society for Mass Spectrometry* **2002**, 13, 284; b)Y. Huang, E. D. Dodds, *Analytical Chemistry* **2013**, 85, 9728.
- [56] D. J. Harvey, C. A. Scarff, M. Edgeworth, M. Crispin, C. N. Scanlan, F. Sobott, S. Allman, K. Baruah, L. Pritchard, J. H. Scrivens, *Electrophoresis* **2013**, 34, 2368.
- [57] a)S. Lee, S. J. Valentine, J. P. Reilly, D. E. Clemmer, *International Journal of Mass Spectrometry* **2012**, 309, 161; b)W. Hoffmann, J. Hofmann, K. Pagel, *Journal of the American Society for Mass Spectrometry* **2014**, 1.
- [58] Y. Liu, D. E. Clemmer, *Analytical Chemistry* **1997**, 69, 2504.
- [59] D. S. Lee, C. Wu, H. H. Hill, *Journal of Chromatography A* **1998**, 822, 1.
- [60] H. Li, K. Giles, B. Bendiak, K. Kaplan, W. F. Siems, H. H. Hill, *Analytical Chemistry* **2012**, 84, 3231.
- [61] a)M. Zhu, B. Bendiak, B. Clowers, H. Hill, Jr., *Analytical and Bioanalytical Chemistry* **2009**, 394, 1853; b)H. Li, B. Bendiak, K. Kaplan, E. Davis, W. F. Siems, H. H. Hill Jr, *International Journal of Mass Spectrometry* **2013**, 352, 9; c)H. L. Li, B. Bendiak, W. F. Siems, D. R. Gang, H. H. Hill, *Analytical Chemistry* **2015**, 87, 2228; d)O. Hernandez, S. Isenberg, V. Steinmetz, G. L. Glish, P. Maitre, *Journal of Physical Chemistry A* **2015**, 119, 6057; e)F. Zhu, S. Lee, S. Valentine, J. Reilly, D. Clemmer, *Journal of the American Society for Mass Spectrometry* **2012**, 23, 2158; f)T. Yamagaki, A. Sato, *Analytical Sciences* **2009**, 25, 985.
- [62] S. Y. Vakhrushev, J. Langridge, I. Campuzano, C. Hughes, J. Peter-Katlinic, *Analytical Chemistry* **2008**, 80, 2506.
- [63] H. L. Li, B. Bendiak, W. F. Siems, D. R. Gang, H. H. Hill, *Rapid Communications in Mass Spectrometry* **2013**, 27, 2699.
- [64] H. Li, B. Bendiak, W. F. Siems, D. R. Gang, H. H. Hill, Jr., *Analytical Chemistry* **2013**, 85, 2760.
- [65] M. E. Pettit, B. Harper, M. R. Brantley, T. Solouki, *Analyst* **2015**, 14, 6886.
- [66] B. Clowers, P. Dwivedi, W. Steiner, H. Hill, B. Bendiak, *Journal of the American Society for Mass Spectrometry* **2005**, 16, 660.
- [67] L. S. Fenn, J. A. McLean, *Molecular Biosystems* **2009**, 5, 1298.
- [68] K. Pagel, D. J. Harvey, *Analytical Chemistry* **2013**, 85, 5138.
- [69] Y. T. Huang, E. D. Dodds, *Analytical Chemistry* **2015**, 87, 5664.

- [70] a)A. Guerrero, C. B. Lebrilla, *International Journal of Mass Spectrometry* **2013**, 354–355, 19; b)L. Han, C. Costello, *Journal of the American Society for Mass Spectrometry* **2011**, 22, 997.
- [71] M. M. Gaye, R. Kurulugama, D. E. Clemmer, *Analyst* **2015**, 140, 6922.
- [72] a)N. M. Lareau, J. C. May, J. A. McLean, *The Analyst* **2015**, 140, 3335; b)D. J. Harvey, F. Sobott, M. Crispin, A. Wrobel, C. Bonomelli, S. Vasiljevic, C. N. Scanlan, C. A. Scarff, K. Thalassinou, J. H. Scrivens, *Journal of the American Society for Mass Spectrometry* **2011**, 22, 568.
- [73] M. Plasencia, D. Isailovic, S. Merenbloom, Y. Mechref, D. Clemmer, *Journal of the American Society for Mass Spectrometry* **2008**, 19, 1706.
- [74] W. B. Struwe, J. L. Benesch, D. J. Harvey, K. Pagel, *The Analyst* **2015**, 140, 6799.
- [75] a)I. Capila, R. J. Linhardt, *Angewandte Chemie International Edition* **2002**, 41, 390; b)Merton Bernfield, Martin Götte, Pyong Woo Park, Ofer Reizes, Marilyn L. Fitzgerald, a. John Lincecum, M. Zako, *Annual Review of Biochemistry* **1999**, 68, 729; c)U. LINDAHL, L. THUNBERG, G. BÄCKSTRÖM, J. RIESENFELD, *Biochemical Society Transactions* **1981**, 9, 499; d)Z. Hu, C. Wang, Y. Xiao, N. Sheng, Y. Chen, Y. Xu, L. Zhang, W. Mo, N. Jing, G. Hu, *Journal of Cell Science* **2009**, 122, 1145; e)H. Zhou, S. Roy, E. Cochran, R. Zouaoui, C. L. Chu, J. Duffner, G. Zhao, S. Smith, Z. Galcheva-Gargova, J. Karlgren, N. Dussault, R. Y. Q. Kwan, E. Moy, M. Barnes, A. Long, C. Honan, Y. W. Qi, Z. Shriver, T. Ganguly, B. Schultes, G. Venkataraman, T. K. Kishimoto, *Plos One* **2011**, 6, e21106.
- [76] a)N. S. Gandhi, R. L. Mancera, *Chemical Biology & Drug Design* **2008**, 72, 455; b)H. Lortat-Jacob, A. Grosdidier, A. Imbert, *Proceedings of the National Academy of Sciences* **2002**, 99, 1229; c)P. Gupta, J. McCarthy, C. Verfaillie, *Blood* **1996**, 87, 3229.
- [77] L. Jin, J. P. Abrahams, R. Skinner, M. Petitou, R. N. Pike, R. W. Carrell, *Proceedings of the National Academy of Sciences* **1997**, 94, 14683.
- [78] a)A. K. Powell, E. A. Yates, D. G. Fernig, J. E. Turnbull, *Glycobiology* **2004**, 14, 17R; b)D. L. Rabenstein, *Natural Product Reports* **2002**, 19, 312; c)J. R. Bishop, M. Schuksz, J. D. Esko, *Nature* **2007**, 446, 1030.
- [79] M. Ly, F. E. Leach, T. N. Laremore, T. Toida, I. J. Amster, R. J. Linhardt, *Nat Chem Biol* **2011**, 7, 827.
- [80] a)S. Faham, R. E. Hileman, J. R. Fromm, R. J. Linhardt, D. C. Rees, *Science* **1996**, 271, 1116; b)E. Sisui, S. Tripathy, J.-M. Mallet, P.-A. Driguez, J.-P. Hérault, P. Sizun, J.-M. Herbert, M. Petitou, P. Sinay, *Biochimie* **2003**, 85, 91.
- [81] Y. Seo, M. R. Schenauer, J. A. Leary, *International Journal of Mass Spectrometry* **2011**, 303, 191.
- [82] a)S. Ricard-Blum, O. Féraud, H. Lortat-Jacob, A. Rencurosi, N. Fukai, F. Dkhissi, D. Vittet, A. Imbert, B. R. Olsen, M. van der Rest, *Journal of Biological Chemistry* **2004**, 279, 2927; b)B. Lages, S. S. Stivala, *Biopolymers* **1973**, 12, 127; c)F. Chevalier, J. Angulo, R. Lucas, Pedro M. Nieto, M. Martín-Lomas, *European Journal of Organic Chemistry* **2002**, 2002, 2367; d)R. González-Iglesias, M. a. A. Pajares, C. Ocal, J. Carlos Espinosa, B. Oesch, M. a. Gasset, *Journal of Molecular Biology* **2002**, 319, 527; e)R. F. Parrish, W. R. Fair, *Biochemical Journal* **1981**, 193, 407; f)S. R. Srinivasan, B. Radhakrishnamurthy, G. S. Berenson, *Archives of Biochemistry and Biophysics* **1975**, 170, 334; g)F. Chevalier, R. Lucas, J. Angulo, M. Martín-Lomas, P. M. Nieto, *Carbohydrate Research* **2004**, 339, 975.
- [83] M. R. Schenauer, J. K. Meissen, Y. Seo, J. B. Ames, J. A. Leary, *Analytical Chemistry* **2009**, 81, 10179.
- [84] R. Miller, W. Wei, R. Schwörer, O. Zubkova, P. Tyler, J. Turnbull, J. Leary, *European Journal of Mass Spectrometry* **2015**, 21, 245.
- [85] a)E. Gray, B. Mulloy, T. W. Barrowcliffe, *Thrombosis and Haemostasis* **2008**, 99, 807; b)D. R. Ferro, A. Provasoli, M. Ragazzi, B. Casu, G. Torri, V. Bossennec, B. Perly, P. Sinay, M. Petitou, J. Choay, *Carbohydrate Research* **1990**, 195, 157; c)P. N. Sanderson, T. N. Huckerby, I. A. Nieduszynski, *Biochemical Journal* **1987**, 243, 175.
- [86] Y. Seo, A. Andaya, J. A. Leary, *Analytical Chemistry* **2012**, 84, 2416.
- [87] B. Mulloy, M. J. Forster, *Glycobiology* **2000**, 10, 1147.
- [88] M. Hermannová, A.-M. Iordache, K. Slovák, V. Havlíček, H. Pelantová, K. Lemr, *Journal of Mass Spectrometry* **2015**, 50, 854.

- [89] M. Kailemia, M. Park, D. Kaplan, A. Venot, G.-J. Boons, L. Li, R. Linhardt, I. J. Amster, *Journal of the American Society for Mass Spectrometry* **2014**, 25, 258.
- [90] J. J. Wolff, T. N. Laremore, F. E. Leach, R. J. Linhardt, I. J. Amster, *European journal of mass spectrometry (Chichester, England)* **2009**, 15, 275.
- [91] a)M. S. Motawia, I. Damager, C. E. Olsen, B. L. Møller, S. B. Engelsen, S. Hansen, L. H. Øgendal, R. Bauer, *Biomacromolecules* **2005**, 6, 143; b)V. G. Murphy, B. Zaslow, A. D. French, *Biopolymers* **1975**, 14, 1487.
- [92] K. Nishinari, K. Kohyama, P. A. Williams, G. O. Phillips, W. Burchard, K. Ogino, *Macromolecules* **1991**, 24, 5590.
- [93] a)J.-H. Huang, E. J. Bakx, H. Gruppen, H. A. Schols, *Rapid Communications in Mass Spectrometry* **2013**, 27, 2279; b)K. Kaneshiro, Y. Fukuyama, S. Iwamoto, S. Sekiya, K. Tanaka, *Analytical Chemistry* **2011**, 83, 3663.
- [94] A. G. M. Leijdekkers, J.-H. Huang, E. J. Bakx, H. Gruppen, H. A. Schols, *Carbohydrate Research* **2015**, 404, 1.
- [95] B. Plancot, G. Vanier, F. Maire, M. Bardor, P. Lerouge, J. M. Farrant, J. Moore, A. Driouich, M. Vitré-Gibouin, C. Afonso, C. Loutelier-Bourhis, *Rapid Communications in Mass Spectrometry* **2014**, 28, 908.
- [96] M. R. Schenauer, J. A. Leary, *International Journal of Mass Spectrometry* **2009**, 287, 70.
- [97] Y. Seo, A. Andaya, C. Bleiholder, J. A. Leary, *Journal of the American Chemical Society* **2013**, 135, 4325.
- [98] Y. Tian, L. Han, A. C. Buckner, B. T. Ruotolo, *Analytical Chemistry* **2015**, 87, 11509.
- [99] B. J. McCullough, J. M. Kalapothakis, W. Chin, K. Taylor, D. J. Clarke, H. Eastwood, D. Campopiano, D. MacMillan, J. Dorin, P. E. Barran, *Physical Chemistry Chemical Physics* **2010**, 12, 3589.
- [100] S. R. Harvey, C. E. MacPhee, B. F. Volkman, P. E. Barran, *Chemical Communications* **2016**.
- [101] H. Li, B. Bendiak, W. Siems, D. Gang, H. Hill, Jr., *International Journal for Ion Mobility Spectrometry* **2013**, 16, 105.
- [102] A. J. Creese, H. J. Cooper, *Analytical Chemistry* **2012**, 84, 2597.
- [103] G. N. Ulasi, A. J. Creese, S. X. Hui, C. W. Penn, H. J. Cooper, *Proteomics* **2015**, 15, 2733.
- [104] K. Brandenburg, O. Holst, in *eLS*, John Wiley & Sons, Ltd, **2001**.
- [105] a)J. B. Robbins, R. Schneerson, *New England Journal of Medicine* **1990**, 323, 1415; b)B. D. Gessner, R. A. Adegbola, *Vaccine* **2008**, 26, Supplement 2, B3.
- [106] E. Miller, D. Salisbury, M. Ramsay, *Vaccine* **2001**, 20, Supplement 1, S58.
- [107] a)J. T. Poolman, *Expert Review of Vaccines* **2004**, 3, 597; b)S. P. Lockhart, J. G. Hackell, B. Fritzell, *Expert Review of Vaccines* **2006**, 5, 553.
- [108] a)A. H. Merrill Jr, M. D. Wang, M. Park, M. C. Sullards, *Trends in Biochemical Sciences* **2007**, 32, 457; b)A. H. Merrill, T. H. Stokes, A. Momin, H. Park, B. J. Portz, S. Kelly, E. Wang, M. C. Sullards, M. D. Wang, *Journal of Lipid Research* **2009**, 50, S97.
- [109] S. Sonnino, V. Chigorno, *Biochimica et Biophysica Acta (BBA) - Reviews on Biomembranes* **2000**, 1469, 63.
- [110] a)R. Yu, Y.-T. Tsai, T. Ariga, *Neurochemical Research* **2012**, 37, 1230; b)E. Posse de Chaves, S. Sipione, *FEBS Letters*, 584, 1748; c)G. van Echten-Deckert, J. Walter, *Progress in Lipid Research* **2012**, 51, 378; d)T. Ariga, M. P. McDonald, R. K. Yu, *Journal of Lipid Research* **2008**, 49, 1157; e)T. Kolter, K. Sandhoff, *Annual Review of Cell and Developmental Biology* **2005**, 21, 81; f)X. Zhang, F. L. Kiechle, *Annals of Clinical & Laboratory Science* **2004**, 34, 3.
- [111] M. R. Wenk, *Nat Rev Drug Discov* **2005**, 4, 594.
- [112] J. Li, R. W. Purves, J. C. Richards, *Analytical Chemistry* **2004**, 76, 4676.
- [113] S. N. Jackson, M. Ugarov, T. Egan, J. D. Post, D. Langlais, J. Albert Schultz, A. S. Woods, *Journal of Mass Spectrometry* **2007**, 42, 1093.
- [114] S. N. Jackson, B. Colsch, T. Egan, E. K. Lewis, J. A. Schultz, A. S. Woods, *Analyst* **2011**, 136, 463.
- [115] S. N. Jackson, D. Barbacci, T. Egan, E. K. Lewis, J. A. Schultz, A. S. Woods, *Analytical Methods* **2014**, 6, 5001.

- [116] S. Jackson, M. Ugarov, J. Post, T. Egan, D. Langlais, J. A. Schultz, A. Woods, *Journal of the American Society for Mass Spectrometry* **2008**, *19*, 1655.
- [117] a)Y. Mechref, Y. Hu, A. Garcia, A. Hussein, *Electrophoresis* **2012**, *33*, 1755; b)H. Liu, N. Zhang, D. Wan, M. Cui, Z. Liu, S. Liu, *Clinical Proteomics* **2014**, *11*, 1.
- [118] D. Isailovic, R. T. Kurulugama, M. D. Plasencia, S. T. Stokes, Z. Kyselova, R. Goldman, Y. Mechref, M. V. Novotny, D. E. Clemmer, *Journal of Proteome Research* **2008**, *7*, 1109.
- [119] A. A. Shvartsburg, W. F. Danielson, R. D. Smith, *Analytical Chemistry* **2010**, *82*, 2456.
- [120] a)S. Lee, M. A. Ewing, F. M. Nachtigall, R. T. Kurulugama, S. J. Valentine, D. E. Clemmer, *Journal of Physical Chemistry B* **2010**, *114*, 12406; b)S. J. Valentine, R. T. Kurulugama, D. E. Clemmer, *Journal of the American Society for Mass Spectrometry* **2011**, *22*, 804; c)R. T. Kurulugama, F. M. Nachtigall, S. J. Valentine, D. E. Clemmer, *Journal of the American Society for Mass Spectrometry* **2011**, *22*, 2049; d)M. A. Ewing, S. M. Zucker, S. J. Valentine, D. E. Clemmer, *Journal of the American Society for Mass Spectrometry* **2013**, *24*, 615; e)M. A. Ewing, C. R. P. Conant, S. M. Zucker, K. J. Griffith, D. E. Clemmer, *Analytical Chemistry* **2015**, *87*, 5132; f)S. Valentine, S. Stokes, R. Kurulugama, F. Nachtigall, D. Clemmer, *Journal of the American Society for Mass Spectrometry* **2009**, *20*, 738.
- [121] a)K. Giles, J. Wildgoose, S. Pringle, J. Garside, P. Carney, P. Nixon, D. Langridge, in *62nd ASMS Conference*, Baltimore, **2014**; b)S. I. Merenbloom, R. S. Glaskin, Z. B. Henson, D. E. Clemmer, *Analytical Chemistry* **2009**, *81*, 1482; c)R. S. Glaskin, M. A. Ewing, D. E. Clemmer, *Analytical Chemistry* **2013**, *85*, 7003.
- [122] L. Cipolla, F. Peri, C. Airoidi, *Anti-Cancer Agents in Medicinal Chemistry* **2008**, *8*, 92.
- [123] C. Jones, *Anais da Academia Brasileira de Ciências* **2005**, *77*, 293.
- [124] J. R. Mascola, G. Stiegler, T. C. VanCott, H. Katinger, C. B. Carpenter, C. E. Hanson, H. Beary, D. Hayes, S. S. Frankel, D. L. Birx, M. G. Lewis, *Nat Med* **2000**, *6*, 207.
- [125] R. D. Astronomo, D. R. Burton, *Nat Rev Drug Discov* **2010**, *9*, 308.
- [126] D. J. Harvey, M. Crispin, C. Bonomelli, J. H. Scrivens, *Journal of the American Society for Mass Spectrometry* **2015**, *26*, 1754.

***Conflict of Interest form**

[Click here to download Conflict of Interest form: coi_disclosure final.pdf](#)